Detection of CHEK2 Ile157Thr Mutation in Cancer Patients by using Allele Specific PCR

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ABSTRACT

Aims: The checkpoint kinase 2 (CHEK2) was identified as a gene in cell cycle control upon DNA damage. The mutation in this gene is associated with some kinds of cancer including breast and lung tumors. The aim of this study was to investigate, I157T missense mutation of CHEK2 gene by Allele specific PCR (ASP) technique in clinical samples of breast and non-small cell lung cancer (NSCLC) samples.

Method: In this study, we investigated I157T mutation of CHEK2 gene in whole blood DNA from 30 breast cancers (BCs), 20 NSCLC patients and 30 control samples. The primers of ASP were designed to detect non-mutant status at the position of 430 CHEK2. The data were analyzed statistically.

Results: The results of the amplification reaction in used samples were indicated the accuracy of designed primers. The results of this study showed that in 70% of NSCLC, 33.34% of BC samples and 63.3% of control samples, the CHEK2-encoding gene contained mutations. Sequencing also confirmed these results. Statistical analysis showed that there was a direct relationship between the change in the position of 157 CHEK2 genes and metastatic NSCLC samples.

Conclusion: In previous studies, mutations at the gene coding for CHEK2 in different cancers have been proven. In the present study, it was found that the designed ASP technique was able to correctly detect the wild-type status in the studied samples and probably could be used to cancer diagnostic analysis, although more samples should be considered.

Key words: Cancer, CHEK2 gene, ASP

INTRODUCTION

In the multistage theory of carcinogenesis, the accumulation of genetic changes leads to the formation of a tumor. Genes involved in the incidence of cancer are classified into several groups, including proto-oncogenes, tumor suppressor genes, genes involved in genome stability and cell migration. In general, many proteins involved in DNA damage repair including BRCA1, Her2 tyrosine kinase receptor (Erbb2), P53, PTEN, MYC, CCND1 etc. Genetic changes in each of these genes have been identified in BC and NSCLC. However, in addition to somatic and hereditary mutations, biological factors can also increase the risk for developing BC and NSCLC [1-3].

At higher levels of DNA damage, mismatch repair (MMR) pathway plays a major role in DNA repairing process and cell-cycle checkpoint control. Cell cycle regulators responding to DNA damage arrest replication and cell cycle. One of these checkpoints of the cell cycle is checkpoint kinase 2 (CHEK2) that acts during MMR repair pathway. CHEK2 plays an imperative role in response to damage or stress induction [4]. The serine/threonine kinase coded by CHEK2 is a tumor suppressor. This protein contains forkhead domains that responding to DNA damage reacts rapidly with phosphorylation and leads to inhibition of CDC25C phosphatase, and ultimately prevents to mitosis which halts the cell cycle in the G1 phase. The mutation in the CHEK2 gene (Homo sapiens checkpoint kinase 2 ID: 1120) is associated with a variety of cancers, including breast, lung, colorectal, brain tumors, etc. [5].

In a study in Denmark in 2006, it was found that individuals harboring mutations in genes known as anticancer barriers including the CHEK2, P53, BRCA1, BRCA2 and ATM genes, are more sensitive to the development of cancer [6].

Bower et al. also identified a pattern of genes controlling the cell cycle associated with luminal B and basal-like cell lines [7].
Also it has been reported that Single nucleotide polymorphism (SNPs) in MMR genes can alter capacity of DNA damage in cancers and modify the function of involved genes [8]. Types of polymorphisms associated with various types of cancer, including Rs178799611, rs38416892, rs17879961, have been reported in CHEK2 gene, which have been investigated by different molecular methods.

However, there are approaches for the detection of polymorphisms. In 2011, Cybulski et al. in Poland examined the risk of prostate cancer by analysis of CHEK2 gene polymorphisms using RFLP and sequencing method [9].

In molecular biology, Allele-Specific PCR technique allows efficient SNP Genotyping in a single reaction of polymerase chain reaction using thermocycler machine. In this method a reverse primer and two forward allele-specific primers amplify different allele-specific products. These fragments are detected by electrophoresis. Evaluation of different molecular methods can help to better diagnose and effectively treatment of cancer.

In this study, the ATT>ACT (nucleotide number 430) mutation associated with amino acid No. 157 (rs17879961) of CHEK2 was studied in clinical samples of breast cancer and NSCLC by the allele specific method.

MATERIALS AND METHOD

Sample preparation

In present case-control study, the whole blood samples were collected from Khansarinejad hospital, Arak, Iran (2017-2018). 50 patient's samples with BC and NSCLC in addition 30 normal samples were used. DNAs were extracted using special kit (Diatom, Iran). Quality of extracted DNA was determined by 1% agarose gel (Genefanavaran, Iran) and the absorbance of A260/A280 nm in spectrophotometer (Eppendorf, Germany).

Allele specific PCR (ASP)

The sequences of designed primers for ASP technique were CHEK2-F: 5’-ATATCCAGCTCCTCTACCAG-3’ and CHEK2-R: 5’-GTGATCTTCTATGTATGGAA-3’. These primers in wild-type status amplified a 375 bp fragment. We used 40 ng genomics DNA, 7.5 μl 2X master-mix (YTA, Iran) and 20 pmol of aforementioned primers in total volume 15 μl per amplification tube.

This reaction was performed in thermocycler system (Eppendorf, Germany) in a touchdown protocol (annealing temperature 62°C-52°C). Results of ASP were analyzed by 1.3% agarose gel in gel doc system (Quantum ST4, Germany).

Sequencing

The confirmation of ASP method, were performed by sequencing. Amplicons were sent to Macrogen Company (South Korea) for sequencing by ABI Applied Biosystems 3730xl system.

Statistics analysis

Results were analyzed using GraphPad 7.00 and SPSS 16.0 software. Medcalc software was used for CI and Odd ratio calculation.

RESULTS

In present study an ASP technique were designed for detection of mutation at ile157Thr hot-spot position of CHEK2 gene.

Figure 1 showed the alignment result of primers for detection of aforementioned missense mutation in used ASP.

Figure 1: (A, B) Chromosomal position of CHEK2 gene in UCSC genome browser and NCBI genebank; (C) Binding sites of primers of allele specific in Mega4 software. Hotspot site of mutation has been shown by star; (D) Schematics view of designed allele specific PCR in present study, in present and absence of mutation at ile157Thr hot-spot position of CHECK2 gene (650 bp and 375 bp respectively)

Figure 2 showed the electrophoresis results of amplification products. These bands indicated the accuracy of the genomic DNA extraction, purification process and the PCR reaction.

Figure 2: Electrophoresis result of different patterns of allele amplification in used samples

The length of PCR Products in wild-type state was 375 bp which are related to TT genotypes. In mutant states is created a non-specific amplification at 650 bp (CC). Also 375 bp and 650 bp indicated TC genotype.

The results of ASP were validated by sequencing. The results showed the rs17879961 genotypes distributions; TT homozygote, TC heterozygote and CC mutant
homozygote variant in cancer group (BC and NSCLC) were different with normal samples (Table 1).

Table 1: The different created patterns by ASP method in normal, breast cancer (BC) and non-small cell lung cancer (NSCLC) samples

<table>
<thead>
<tr>
<th>Genotype</th>
<th>BC (%) n=30</th>
<th>Control (%) n=30</th>
<th>95% CI</th>
<th>Odds ratio</th>
<th>z statistic</th>
<th>Significance level</th>
</tr>
</thead>
<tbody>
<tr>
<td>TC heterozygote</td>
<td>6.66</td>
<td>26.66</td>
<td>0.0839 to 0.4936</td>
<td>0.2035</td>
<td>3.522</td>
<td>p=0.0004</td>
</tr>
<tr>
<td>CC homozygote</td>
<td>33.33</td>
<td>63.33</td>
<td>0.1474 to 0.4794</td>
<td>0.2658</td>
<td>4.404</td>
<td>p=0.0001</td>
</tr>
<tr>
<td>TT homozygote (WT)</td>
<td>60</td>
<td>10</td>
<td>6.2751 to 29.0434</td>
<td>13.5</td>
<td>6.659</td>
<td>p=0.0001</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Genotype</th>
<th>NSCLC n=20</th>
<th>Control (%) n=30</th>
<th>95% CI</th>
<th>Odds ratio</th>
<th>z statistic</th>
<th>Significance level</th>
</tr>
</thead>
<tbody>
<tr>
<td>TC heterozygote</td>
<td>0</td>
<td>26.66</td>
<td>0.0007981 to 0.2215</td>
<td>0.0133</td>
<td>3.01</td>
<td>p=0.0026</td>
</tr>
<tr>
<td>CC homozygote</td>
<td>70</td>
<td>63.33</td>
<td>0.7599 to 2.4714</td>
<td>1.3704</td>
<td>1.047</td>
<td>p=0.2950</td>
</tr>
<tr>
<td>TT homozygote (WT)</td>
<td>30</td>
<td>10</td>
<td>1.7666 to 8.4217</td>
<td>3.0571</td>
<td>3.388</td>
<td>p=0.0007</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Genotype</th>
<th>NSCLC n=20</th>
<th>BC (%) n=30</th>
<th>95% CI</th>
<th>Odds ratio</th>
<th>z statistic</th>
<th>Significance level</th>
</tr>
</thead>
<tbody>
<tr>
<td>TC heterozygote</td>
<td>0</td>
<td>6.66</td>
<td>0.003494 to 1.1011</td>
<td>0.06202</td>
<td>1.894</td>
<td>p=0.0582</td>
</tr>
<tr>
<td>CC homozygote</td>
<td>70</td>
<td>33.33</td>
<td>2.4848 to 8.2412</td>
<td>4.5253</td>
<td>4.936</td>
<td>p&lt;0.0001</td>
</tr>
<tr>
<td>TT homozygote (WT)</td>
<td>30</td>
<td>60</td>
<td>1.9486 to 6.2866</td>
<td>3.5</td>
<td>4.193</td>
<td>p=0.0001</td>
</tr>
</tbody>
</table>

DISCUSSION

CHEK2 is one of the genes involved in various types of lung cancer, as well as classified as one of the 10 altered genes associated with hereditary BC [10-12]. In addition to, CHEK2 gene expression has been observed in colorectal, ovarian, brain and prostate cancers [13-15]. Processing of DNA damage via MMR pathway, CHEK2 nuclear protein interacts with BRCA1 and phosphorylates it and returned to survival after DNA damage. CHEK2 might be a target gene in the treatment of patients with lung cancer, however protective effect of mutant CHEK2 alleles is not yet known [16].

The ATM/CHK2 pathway regulates the checkpoint of G1 phase in cell cycle, through activation of the p53 that prevents cells from entering S-phase [17]. Several SNPS in the encoding region of this gene have been studied in previous studies. In the present study, we developed an in-house method for detecting the mutation in the rs17879961 of CHEK2 gene, which leads to nucleotide conversion T to C and a changing in the second nucleotide of the codon 157 (isoleucine acid to threonine amino acid). The cost of used ASP method was very low and feasible in our molecular lab.

In 2016, the association between CHEK2 polymorphism and platinum-based chemotherapy efficiency in Chinese non-smokers with NSCLC was investigated and revealed that individuals with rs4035540 A/G genotype more survived [18].

In another study in 2018, a variety of polymorphisms in the CHEK2 gene, including rs1547014 and rs738722 were detected, which could possibly predict a HBV-associated hepatocellular carcinoma patient’s response to therapy [19]. In a study in 2018 in the United States, CHEK2 SNPs and ATM variants were also examined [20]. In our study, we analyzed lung and BC cancer samples, however total number with CC genotypes in lung cancer were more than BC samples (Table 1). In a 2008 study in China, the association of 1100delC 1111C>T mutation with susceptibility to BC was determined [21]. Their study showed that CHEK2 1100delC mutation may be a rare variant in Chinese populations and may not contribute to predisposition for BC. However, there may be an association between BC genetic susceptibility in China and this variant.

In our study, some of the demographic data (Table 2) of cancer patients with mutation in the amino acid 157 of the CHECK2 gene were examined and elucidated that number of metastatic and non-metastatic of NSCLC samples have alteration of 157 codon of CHEK2 gene. Although, most of the BC samples in this study were non-metastatic and among them, 60% don’t have mutation in the codon 157 of CHEK2 gene.

Table 2: Demographic data of cancer patients with mutation in the amino acid 157 of the CHECK2 gene

<table>
<thead>
<tr>
<th>Characteristic-NSCLC</th>
<th>Detail</th>
<th>No. of patients (%)</th>
<th>Characteristic-BC</th>
<th>Detail</th>
<th>No. of patients (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Median age, range</td>
<td>≤ 58</td>
<td>48</td>
<td>Age</td>
<td>&lt;40</td>
<td>23</td>
</tr>
<tr>
<td></td>
<td>&gt;58</td>
<td>52</td>
<td>&gt;40</td>
<td>77</td>
<td></td>
</tr>
<tr>
<td>Gender</td>
<td>Women</td>
<td>20</td>
<td>Menopausal age</td>
<td>No</td>
<td>48</td>
</tr>
<tr>
<td></td>
<td>Men</td>
<td>80</td>
<td>&lt;52</td>
<td>8</td>
<td></td>
</tr>
</tbody>
</table>
Luo et al., investigated the mechanism of change in the function of the CHEK2 in triple Negative BC cells, chemotherapy-resistant cell line. In their study, it was found that in the MDA-MB-231 cell lines, Y390C mutation leads to resistance to chemotherapy through the effect on the cell cycle arrest by activating the p53 and CHEK2 gene in the apoptosis process [22].

Also aforementioned mutation disrupted the normal activity of the CHEK2 gene in patients with high risk of BC. CHEK2 alteration is related to CRrispr-cas9 istop, new attractive subjects in molecular genetics also [23]. Mutations in the CHEK2 gene including I157T have been reported in the different cancers. However due to change in second nucleotide which converts isoleucine to threonine amino acids, that both of these amino acids belonged to same family with similar chemical properties.

CONCLUSION

In present study, we detected I157T mutation in CHECK2 gene using an ASP technique. We confirmed this alteration of CHECK2 gene is related to metastatic status of NSCLC, therefore, could be used as a marker of metastatic probably. Although the larger sample size of cancer patients will need to accurate analysis.

ACKNOWLEDGMENT

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CONFLICT OF INTEREST

The authors declare that there is no conflict of interest regarding the publication of this manuscript.

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