

Emerging Role of the Neuronal Sweet Taste Receptor Heterodimer, T1R2+T1R3, in Cognitive Functioning

Menizibeya O Welcome*

Department of Physiology, College of Health Sciences, The Nile University of Nigeria, Nigeria

ABSTRACT

Diseases involving cognitive impairments present substantial burden to sufferers, families, caregivers and public health. Unfortunately, however, the mechanisms underlying cognitive disorders are yet to be fully unraveled. Taste receptors, initially discovered in the oral cavity, are now believed to be ubiquitously expressed in cells and tissues of the body, including central neurons. Recent investigations have shown that the sweet taste receptor heterodimer, T1R2+T1R3, plays a crucial role in cognitive functioning, suggesting that dysfunctions in sweet taste receptor signaling may underlie cognitive impairment in some brain pathologies. In this mini review, I integrate very recent data that suggest possible molecular mechanisms, linking neuronal sweet taste sensing to cognitive functioning. Future research directions and targeting of sweet taste receptor for potential treatment of brain diseases, involving sweet taste receptor dysfunctions are also discussed.

Key words: T1R2+T1R3, Sweet taste receptors, Neuron, Cognition, Cognitive impairment

HOW TO CITE THIS ARTICLE: Menizibeya O Welcome*, Emerging role of the neuronal sweet taste receptor heterodimer, T1R2+T1R3, in cognitive functioning, J Res Med Dent Sci, 2018, 6 (5):264-272

Corresponding author: Menizibeya 0 Welcome e-mail⊠: welcome.menizibeya@nileuniversity.edu.ng Received: 03/09/2018 Accepted: 23/10/2018

INTRODUCTION

The word "cognition" refers to a variety of subcomponents of higher brain functions such as thinking, reasoning, planning, attention, memory, learning, which are executed by the human brain [1]. However, depending on the context, cognitive functioning can simply refer to learning and memory [2]. Different definitions of cognitive function have been used by other authors [3–11]. Cognitive function is crucial to the effectiveness and ongoing daily life activities of humans [1]. Cognitive disorder or impairment, defined as cognitive decline greater than expected for the person's age and education level, which may interfere with the professional or social life of the sufferer [12], is believed to be the major determinant of disability in late life [1]. Cognitive impairment is a syndrome of several diseases, including Alzheimer's, Parkinson's, and Huntington disease, and multiple sclerosis [12-17]. But Alzheimer's disease accounts for a higher incidence of cognitive impairment: about 60-75% of all cases of cognitive impairment or dementia (severe cognitive impairment) worldwide [18-20]. It should be mentioned that cognitive impairment is a serious problem that can develop even in children and adolescents [21-23]. Depending on the degree of severity, cognitive impairment observed in different neurological diseases poses enormous financial burden

to sufferers, families, caregivers and public health [24–28]. Due to the growing prevalence and associated mortality of the neurological diseases, associated with cognitive impairment [24,26], it is evidently necessary to search for new frontiers that will lead to potentially new treatment of cognitive impairment or diseases that predispose to cognitive decline.

Although the mechanisms of development of cognitive decline in different neurological diseases differ, research indicates that neuro- and glio-inflammatory reactions and cell death are critical in almost all diseases associated with cognitive impairment [29,30]. Very recently, emerging data have revealed that dysfunctions of sweet taste receptor signaling may be responsible for cognitive impairment [31], confirming our initial hypothesis about the role of sweet taste receptors in regulation of cognitive functioning [32]. Surprisingly, dysfunctions in sweet taste receptor signaling were also associated with inflammatory response pathway [31]. Zhou et al. recently showed that sweet taste receptors function as pivotal immune sentinels. The authors revealed that key components of the taste signaling cascades (α -gustducin; phospholipase C β 2, PLC₆₂; monovalent selective cation channel, TRPM5) were down regulated together with abnormal increase in glucose level, which in turn predisposed to elevated intracellular reactive oxygen species, decrease Ca2+ and subsequent activation of NLRP3 (NLR Family Pyrin 3) domain containing inflammasome signaling, possibly mediated via NF-κB (nuclear factor κB)

activation [31]. Indeed the role of sweet taste receptor signaling in immunity is an important mechanism that link sweet taste receptor with neurodegenerative diseases and glucose metabolism [31]. Emerging data indicate that neuro- and glio-inflammation represents the underlying mechanisms of the etiopathogensis of neurodegenerative diseases [33,34]. Moreover, previous studies have reported not only taste dysfunctions, but also astrocyte-neuron glucose metabolic dysfunctions in neurodegenerative diseases such as multiple sclerosis, Alzheimer's, and Parkinson's diseases [35–41].

Sweet taste receptors are grouped as type 1 (T1R). Functional sweet taste receptors mainly function as dimers and are activated by a plethora of natural and artificial ligands. The natural ligands for sweet taste receptors include monosaccharides (glucose, galactose, fructose), disaccharides (sucrose, maltose, lactose), amino acids (glycine, alanine, threonine, D-histidine, D-tryptophan), and sweet proteins (brazzein, thaumatin, monellin). The artificial ligands for sweet taste receptors include acesulfame potassium, aspartame, cyclamate, sucralose saccharin, glycyrrhizin, and neotame [32]. The second (T2R) type of taste receptors is responsible for sensing bitter substances. This review is concerned only with type 1 taste receptors, in particular, the taste receptor T1R2+T1R3 heterodimer. Taste receptors were initially discovered in the oral cavity, then in other regions of the gastrointestinal tract [42-45], and later in endocrine pancreatic cells [46]. Taste receptors have been found in blood cells, in particular, macrophages [47], and respiratory track [48] and are now known to be ubiquitously expressed in the body. Brain (neuronal) type T1R was fairly recently discovered [49]. Importantly, the authors [49] identified the principal (functional) taste receptor T1R2+T1R3 heterodimer, and all subtypes of sweet taste receptors (T1R1, T1R2 and T1R3) in the neurons of hypothalamus, CA area and dentate gyrus of the hippocampus, and cortex [49]. Interestingly, these brain regions are implicated in both energy homeostasis and cognitive functioning [32]. Indeed discoverers of neuronal sweet taste receptors also showed that these receptors are essential for controlling cerebral metabolism. In a relatively recent review [50], our research group has suggested that T1R2+T1R3 heterodimer is the master coordinator of the astrocyte-neuron metabolic machinery, and highlighted the molecular pathways that link taste sensing with metabolism. Truly, the relationship between glucose metabolism and cognitive functioning has been reported in a couple of studies [51-54]. So, extra-oral sweet taste receptors can mediate a variety of physiological processes, including metabolism, inflammatory responses, and a range of cellular activities [48,55,56].

In light of accumulating evidences, suggesting that sweet taste receptors play an integral role in cognitive functioning and that dysfunctions in components of the signaling pathway of sweet taste receptor may underlie cognitive disorders in some brain pathologies, in this mini review, I integrate very recent studies that suggest possible molecular mechanisms, linking T1R2+T1R3 sweet taste receptor heterodimer sensing in neurons with cognitive functioning. This review also discusses potential research directions that may yield new lines of treatment for brain disorders, involving cognitive impairment such as Alzheimer's disease.

Signal transduction pathway of T1R2+T1R3 receptor heterodimer

The mechanisms of taste receptor signaling involve activation by sweet substance (e.g. glucose), resulting to downstream signaling that culminate in stimulation



Figure 1: Signal transduction pathway of T1R2+T1R3 receptor

of neuropeptide secretion, activation of transcription factors and gene expression (Figure 1).

In Figure 1, Neuronal T1R2+T1R3 activation by glucose induces intracellular signaling in which the α -subunit gustducin dissociates from the $\beta\gamma$ subunits upon exchange of GDP for GTP [57–59]. The α -gustducin activates the membrane bound enzyme adenylate cyclase, which mediates the conversion of ATP to cAMP [60]. However, depending on the isoforms, α -gustducin can also activate phosphodiesterase (PDE) to decrease intracellular cAMP levels [59-61]. Increase in cAMP level can stimulate cAMP-dependent ion channels and protein kinases such as PKA, which when activated, recruits secretory granules towards the plasma membrane for exocytosis. Activated PKA can also translocate to the nucleus to activate gene expression [60]. The $\beta\gamma$ subunits activate phospholipase $C\beta$ (PLC β) with formation of 1,4,5-inositol trisphosphate (IP3). IP3 is responsible for the release of Ca²⁺ from intracellular stores. Increased Ca²⁺ level in the cytosol activates calcium dependent kinases, monovalent selective cation channel (TRPM5), calcium homeostasis modulator 1 (CALHM1) and other receptors [59,61–63]. The evoked increase in Ca^{2+} may also lead to release of ATP via dedicated ion channel receptor [64]. The voltage-gated ion channel, CALHM1, has been implicated in taste related ATP release [43]. DAG, released upon βy subunit activation, can stimulate protein kinase C (PKC), which phosphorylates or activates several intracellular targets, including transcription factors. A couple of secretory neuro-peptides and hormones may be released in T1R2+T1R3 receptor downstream signaling [32]. Events resulting to activation of ion channels can lead to membrane depolarization and subsequent generation of action potential [64]. Taste receptor T1R2+T1R3 also cooperates with the membrane glucosensors (e.g. GLUT2) to control glucose uptake. Gap junctions, hemichannels, and pannexins play a role in homeostasis of ions and other signaling molecules in many physiological processes occurring in the neuron [32].

Deciphering the mechanisms of neuronal T1R2+T1R3 involvement in cognitive functioning

Cognition is a higher mental function of the neocortex, required for almost all activities of humans as well as survival. Cognition depends not only on neocortical, but also hippocampal processes. This higher brain function is also related to several facets of behavior and emotion [32,65]. Based on accumulating data, it is suggested in this paper that T1R2+T1R3 signaling is coupled to neural network of cognition *via* neurotransmitter, cAMP-, Ca²⁺-dependent and PKC related transcription activation and the expression of early response gene (also known as immediate early genes). Indeed cAMP-, Ca²⁺-dependent signaling and activity of certain protein kinases (including PKC and PKA) that culminate in activation of transcription of early response genes have been known to underlie memory formation, storage

and retrieval. Early response genes are a class of genes that increase transiently in expression in response to extracellular signals such as neurotransmitters. Early response genes code for transcription factors that transiently increase transcription in certain areas of the brain upon stimulation. The transcription factors play a crucial role in signal transduction cascades, necessary for memory formation and consolidation [66]. They form important molecular nexus between neuronal activity and cognition [67].

T1R2+T1R3 signaling is coupled to cognitive network *via* stimulation of neurotransmitter secretion, transcription signaling and early gene responses

From the discussion above, it is evident that cognitive functioning involves interaction of multiple players at different levels: neuromodulators, neurohormones, neurotransmitters, energy substrates (mainly glucose), transcription factors [68-70]. Transmitter and molecules involved in cognitive functioning include but are not limited to acetylcholine, serotonin, dopamine, d-serine, ATP and glutamate [32,68,71]. Recall that ATP and glutamate are crucial neurotransmitters, involved in sweet taste receptor signaling (Figure 1). Therefore, sweet taste signaling, at least via ATP and glutamate, can substantially affect cognitive functioning. Emerging research has shown that taste receptors stimulate the secretion of not only ATP and glutamate, but also serotonin and dopamine [72,73]. These transmitter molecules and a range of receptors (including N-methyl-D-aspartate, NMDA receptors; α-amino-3-hydroxy-5methyl-4-isoxazolepropionic, AMPA, receptor) [74] mediate the induction of long-term potentiation (LTP), and synaptic plasticity. Importantly, LTP and synaptic plasticity are factors that are essential for storage and retrieval of neural information [32,68]. LTP and synaptic plasticity are associated with activity dependent neuronal signaling and activity of transcription factors, and are important for memory consolidation [59-61,75,76]. For example, stimulation of group I metabotropic glutamate receptors, G_a coupled receptor (mGluR1 and mGluR5) by glutamate can trigger a range of cellular signaling (PLCβ, certain isoforms of adenylate cyclase and MAP kinase) that ultimately result in LTP induction that underlie long-term synaptic plasticity and metaplasticity in neuronal assemblies [68,76,77-79]. Several protein kinases including MAP kinase, CaMKIIa, CaMKB, PKA and PKC are involved in LTP induction and synaptic plasticity [80]. These protein kinases, in addition to cAMP and calcium ions, can regulate the activities of transcription factors such as enhancer-binding protein (EBP), CREB (cAMP related element binding protein), early growth response protein (egr) (also known as zinc finger protein 225, Zif268; nerve growth factor-induced protein A, NGFI-A), activator protein-1 (AP-1), Rel/NFκB, Elk-1, c-Jun, Jun-B, Jun-D, c-fos [32,66,81-84] and neuronal PAS domain protein 4 (Npas4) [67], which are implicated in long-term synaptic plasticity and memory consolidation. The signaling of these transcription factors is largely dependent on neuronal activity [74,83]. It is believed that some of these transcription factors also influence_activity-dependent expression of cytoskeleton-associated protein (e.g. activity-regulated cytoskeletal-associated protein, Arc; and cofflin) [85,86] as well as girdin (girders of actin filament) [87] to modulate a range of physiological processes associated with synaptic plasticity and metaplasticity. Interestingly, Ca²⁺, cAMP, PKA, and MAP kinase activation have been shown to be involved in Arc gene induction [88]. The cytoskeletal protein, Arc, acts as a master regulator of synaptic plasticity and long-term memory formation *via* regulation by MAP kinase phosphorylation [89].

The results of some researchers indicate that late response factors such as brain-derived neurotrophic factor (BDNF) play a pivotal role in LTP induction and synaptic plasticity to influence memory formation, storage and retrieval [90].

Finally, it should mentioned that the expression of memory related genes are dependent on increased recruitment of histone deacetylase 2, an epigenetic enzyme, which forms a repressor complex with zinc finger transcription factor that regulates cognitive functioning [84,91,92]. The role of epigenetic enzymes in cognitive functioning has been reviewed elsewhere [93–97].

Cognitive processes are coupled to cerebral glucose metabolism by the T1R2+T1R3 receptor heterodimer

From Figure 2, it is obvious that T1R2+T1R3 signaling is coupled to cognitive processes *via* glucose metabolism.

In addition to the signaling pathway triggered upon activation of taste receptor heterodimer as defined in Figure 1, T1R2+T1R3 can stimulate the functions of GLUT2 to increase glucose uptake. Increase in glucose uptake is associated with cerebral insulin signaling [32]. (Cerebral insulin can signal downstream to regulate the activities of early genes or SIRT1) [98,99]. Glucose, a crucial energy molecule, in the cytosol undergoes several metabolic reactions to produce ATP and other substances required for physiological processes in neurons. Increased level of cellular ATP promotes inhibition of potassium fluxes by the K-ATP channel. The level of ATP in the cell is constantly been monitored by energy sensors, in particularly, AMPK, which also interacts with SIRT1 to regulate the activities of early genes and the epigenome. Again, under the action of adenylate cyclase, ATP can be converted to cAMP, which in turn activates PKA. The activated PKA can interact with several effectors: It activates secretory granule exocytosis and transcription factors (Neuron A) [32,98,99]. The exocytosed molecules (e.g. neurohormones) diffuse to neighboring neurons to



Figure 2: Mechanisms of coupling of T1R2+T1R3 signaling to cognitive network of the brain

stimulate their cognate receptors, which trigger a series of cascades, culminating in increased cytosolic calcium, and subsequently, activation of certain membrane receptors, transcription factors, and the expression of early response genes that are involved in synaptic plasticity, memory and learning (Neuron B) [32].

The relationship between glucose metabolism and cognitive processes has been demonstrated in a couple of studies [51,100,101]. This functional cross-talk is possible due to the interaction between metabolic products of glucose and cognitive circuitry [102]. Metabolic signals have been shown to determine reward or emotion [103-106] from sugar ingestion. A recent study by Chan et al. [107] revealed that stimulation of sweet taste receptors by ingestion of non-nutritive, noncalorie sweeteners stimulated glucose transport, incretin effect and insulin secretion. The result was enhancement of glucose metabolism. Similar results were reported by Murovets et al. [108], who showed an association between T1R3 defect and disorder of glucose transport in mice. Furthermore, it was shown that activation of brain sweet taste receptors was associated with incretin effect.

The different substances produced from glucose via series of metabolic processes-glycolysis and oxidative phosphorylations are involved in maintenance of cognitive systems [109]. Accumulating data have shown that metabolic coupling to cognition is believed to occur via lactate synthesis [65,80,110,111]. Neuronal metabolism, culminating in lactate production, is involved in LTP induction, synaptic plasticity, memory formation, which are thought to involve the activation of CREB, Arc and cofilin [112]. Relatively recent animal study demonstrated that defect in lactate transport is associated with disorder in memory formation [112]. Though lactate itself may not directly enhance cognition, its metabolic products in the TCA cycle and their activation of transcription factors may be necessary for lactate enhancement of cognitive functions [112]. Neuron-derived lactate appears to stimulate the expression of genes (Arc, c-Fos, and Zif268), which are associated with synaptic plasticity [113]. Indeed lactate may serve as substrate for the synthesis of synaptic or structural proteins such as Arc and cofflin, which may occur *via* transcription regulation [80]. In this regard, previous data have consistently shown that AMPK, CREB and mTOR are involved in both glucose metabolism and memory functions [71]. Importantly, these molecules are strongly involved in T1R2+T1R3 signaling [32]. Furthermore, Yang et al. [113] reported that infusion of lactate resulted to potentiation of NMDA receptor activity, which is involved in LTP induction and memory formation. Data have consistently shown that AMPK and other metabolic sensors are involved in both glucose metabolism and cognitive functions [114]. Thus disorders involving metabolic sensors such as AMPK and mTOR can result to cognitive decline, and more importantly, have been associated with neuropsychiatric symptoms, including intellectual disability, specific neuropsychological deficits, and other behavioral and cognitive disorders [115].

CONCLUSION

The T1R2+T1R3 heterodimer is a functional sweet taste receptor that drives cerebral glucose metabolism and regulates cognitive functioning. The possible molecular mechanisms, linking neuronal sweet taste sensing to cognitive functioning involves multiple cross-talks with membrane glucosensors (GLUT2, K_{ATP}), cytosolic glucosensors and epigenetic factors (SIRT1, AMPK and mTOR), and transcription factors (EBP, CREB, egr, AP-1, Rel/NF- κ B, Elk-1, c-Jun, Jun-B, Jun-D, c-fos, Npas4), and other signaling molecules: Ca²⁺, cAMP, protein kinases (PKA, PKC, MAP kinase). Activity dependent signaling by neuronal T1R2+T1R3 is critical for both metabolic and cognitive processes in health and disease.

FUTURE DIRECTIONS

Since T1R2+T1R3 heterodimer signaling is related to cognitive functioning (Figure 2) [31,32], future research will investigate the effects of downstream signaling of neuronal sweet taste receptors on cerebral and global glucose homeostasis in health and disease. Specific attention will be given to diabetes mellitus, prediabetes, and brain diseases involving both cognitive decline and metabolic dysregulation (e.g. neurodegenerative disorders-Alzheimer, Parkinson diseases etc.). Further investigations into the effects of different pharmacological agents on sweet taste receptor functions can yield potential treatment options for brain diseases, involving both metabolic and cognitive dysregulation.

CONFLICT OF INTEREST

There is no conflict of interest regarding the publication of this paper.

REFERENCES

- 1. Borson S. Cognition, aging, and disabilities: Conceptual issues. Phys Med Rehabil Clin 2010; 21:375-82.
- 2. Brem AK, Ran K, Pascual-Leone A. Learning and memory. Handb Clin Neurol 2013; 116:693-737.
- 3. Wallin A, Kettunen P, Johansson PM, et al. Cognitive medicine: A new approach in health care science. BMC Psychiatry 2018; 18:42.
- 4. Tacca MC. Commonalities between perception and cognition. Front Psychol 2011; 2:358.
- 5. Jeannerod M, Jacob P. Visual cognition: A new look at the two-visual systems model. Neuropsychol 2005; 43:301-12.
- 6. Piccinini G, Scarantino A. Information processing, computation, and cognition. J Biol

Phys 2011; 37:1-38.

- Mackie MA, Van Dam NT, Fan J. Cognitive control and attentional functions. Brain Cogn 2013; 82:301-12.
- Chaney DW. An overview of the first use of the terms cognition and behavior. Behav Sci 2013; 3:143-53.
- 9. Duncan S, Barrett LF. Affect is a form of cognition: A neurobiological analysis. Cogn Emot 2007; 21:1184-211.
- 10. Perlovsky L. Language and cognition. Neural Netw 2009; 22:247-57.
- Robbins TW. Cognition: The ultimate brain function. Neuropsychopharmacology 2011; 36:1–2.
- 12. Gauthier S, Reisberg B, Zaudig M, et al. Mild cognitive impairment. Lancet 2006; 367:1262-70.
- 13. Bidelman GM, Lowther JE, Tak SH, et al. Mild cognitive impairment is characterized by deficient brainstem and cortical representations of speech. J Neurosci 2017; 3700-16.
- 14. Patti F, Amato MP, Trojano M, et al. Cognitive impairment and its relation with disease measures in mildly disabled patients with relapsing-remitting multiple sclerosis: Baseline results from the cognitive impairment in multiple sclerosis (COGIMUS) study. Mult Scler J 2009; 15:779-88.
- 15. Paulsen JS. Cognitive impairment in Huntington disease: Diagnosis and treatment. Curr Neurol Neurosci Rep 2011; 11:474.
- Goldman JG, Litvan I. Mild cognitive impairment in Parkinson's disease. Minerva Med 2011; 102:441–59.
- 17. Mufson EJ, Binder L, Counts SE, et al. Mild cognitive impairment: Pathology and mechanisms. Acta Neuropathol 2012; 123:13-30.
- Alkasir R, Li J, Li X, et al. Human gut microbiota: The links with dementia development. Protein Cell 2017; 8:90-102.
- 19. Dong S, Duan Y, Hu Y, et al. Advances in the pathogenesis of Alzheimer's disease: A reevaluation of amyloid cascade hypothesis. Transl Neurodegener 2012; 1:18.
- 20. Hung YN, Kadziola Z, Brnabic AJ, et al. The epidemiology and burden of Alzheimer's disease in Taiwan utilizing data from the national health insurance research database. Clinicoecon Outcomes Res 2016; 8:387.
- 21. Julian L, Serafin D, Charvet L, et al. Cognitive impairment occurs in children and adolescents with multiple sclerosis: Results from a United States network. J Child Neurol 2013; 28:102-7.
- 22. Wing L. Language, social, and cognitive impairments in autism and severe mental retardation. J Autism Dev Disord 1981; 11:31-44.
- 23. Breau LM, Camfield CS, McGrath PJ, et al. The incidence of pain in children with severe

cognitive impairments. Arch Pediatr Adolesc Med 2003; 157:1219-26.

- 24. Mavrodaris A, Powell J, Thorogood M. Prevalences of dementia and cognitive impairment among older people in sub-Saharan Africa: A systematic review. Bull World Health Organ 2013; 91:773-83.
- 25. Paradise M, McCade D, Hickie IB, et al. Caregiver burden in mild cognitive impairment. Aging Ment Health 2015; 19:72-8.
- 26. Hugo J, Ganguli M. Dementia and cognitive impairment: Epidemiology, diagnosis, and treatment. Clin Geriatr Med 2014; 30:421-42.
- 27. Jones AJ, Kuijer RG, Livingston L, et al. Caregiver burden is increased in Parkinson's disease with mild cognitive impairment (PD-MCI). Transl Neurodegener 2017; 6:17.
- 28. Werner P. Mild cognitive impairment and caregiver burden: A critical review and research agenda. Public Health Rev 2012; 34:16.
- 29. Halliday GM, Leverenz JB, Schneider JS, et al. The neurobiological basis of cognitive impairment in Parkinson's disease. Mov Disord 2014; 29:634-50.
- 30. Hong H, Kim BS, Im HI. Pathophysiological role of neuroinflammation in neurodegenerative diseases and psychiatric disorders. Int Neurourol J 2016; 20:S2.
- 31. Zhou L, Huang W, Xu Y, et al. Sweet taste receptors mediated ROS-NLRP3 inflammasome signaling activation: Implications for diabetic nephropathy. J Diabetes Res 2018; 2018.
- 32. Welcome MO, Mastorakis NE, Pereverzev VA. Sweet taste receptor signaling network: Possible implication for cognitive functioning. Neurol Res Int 2015; 2015.
- Glass CK, Saijo K, Winner B, et al. Mechanisms underlying inflammation in neurodegeneration. Cell 2010; 140:918-34.
- 34. Ransohoff RM. How neuroinflammation contributes to neurodegeneration. Science 2016; 353:777-83.
- 35. w w w . i n t e c h o p e n . c o m / b o o k s / neurodegenerative-diseases-processesprevention-protection-and-monitoring/role-ofastrocytes-in-neurodegenerative-diseases
- 36. Sidoryk-Wegrzynowicz M, Wegrzynowicz M, Lee E, et al. Role of astrocytes in brain function and disease. Toxicol Pathol 2011; 39:115-23.
- 37. Suzanne M, Tong M. Brain metabolic dysfunction at the core of Alzheimer's disease. Biochem Pharmacol 2014; 88:548-59.
- Bromley SM. Smell and taste disorders: A primary care approach. Am Fam Physician 2000; 61:427-36.
- 39. DeVere R. Disorders of taste and smell. Continuum (Minneap Minn) 2017; 23:421-46.
- 40. Devere R. Smell and taste in clinical neurology-five new things. Neurol Clin Pract 2012; 2:208-14.
- 41. Aliani M, Udenigwe CC, Girgih AT, et al. Aroma

and taste perceptions with Alzheimer disease and stroke. Crit Rev Food Sci Nutr 2013; 53:760-9.

- 42. Adler E, Hoon MA, Mueller KL, et al. A novel family of mammalian taste receptors. Cell 2000; 100:693-702.
- 43. Hoon MA, Adler E, Lindemeier J, et al. Putative mammalian taste receptors: A class of tastespecific GPCRs with distinct topographic selectivity. Cell 1999; 96:541-51.
- 44. Mace OJ, Affleck J, Patel N, et al. Sweet taste receptors in rat small intestine stimulate glucose absorption through apical GLUT2. J Physiol 2007; 582:379-92.
- 45. Jang HJ, Kokrashvili Z, Theodorakis MJ, et al. Gut-expressed gustducin and taste receptors regulate secretion of glucagon-like peptide-1. Proc Natl Acad Sci USA 2007; 104:15069-74.
- 46. Nakagawa Y, Nagasawa M, Yamada S, et al. Sweet taste receptor expressed in pancreatic β -cells activates the calcium and cyclic AMP signaling systems and stimulates insulin secretion. PloS One 2009; 4:e5106.
- 47. Taya K, Hirose K, Hamada S. Trehalose inhibits inflammatory cytokine production by protecting IκB-α reduction in mouse peritoneal macrophages. Arch Oral Biol 2009; 54:749-56.
- 48. Lee RJ, Cohen NA. Taste receptors in innate immunity. Cell Mol Life Sci 2015; 72:217-36.
- 49. Ren X, Zhou L, Terwilliger R, et al. Sweet taste signaling functions as a hypothalamic glucose sensor. Front Integr Neurosci 2009; 3:12.
- 50. Welcome MO, Mastorakis NE. Emerging concepts in brain glucose metabolic functions: From Glucose sensing to how the sweet taste of glucose regulates its own metabolism in astrocytes and neurons. Neuromolecular Med 2018; 1-20.
- 51. Welcome MO, Pereverzev VA. Glycemic allostasis during mental activities on fasting in non alcohol users and alcohol users with different durations of abstinence. Ann Med Health Sci Res 2014; 4:199-207.
- 52. Welcome MO, Pereverzeva EV, Pereverzev VA. A novel psychophysiological model of the effect of alcohol use on academic performance of male medical students of Belarusian State Medical University. Int J Collab Res Intern Med Public Health 2010; 2:183.
- 53. Welcome MO, Dane Ş, Mastorakis NE, et al. Glucoallostasis and higher integrative brain functions, In: Advances in Psychobiology. New York: Nova Science Publishers 2018; 119–236.
- 54. Welcome MO, Mastorakis NE, Pereverzev VA. Multi-level system coupling of error commission, detection and correction in the error monitoring and processing system are required for high precision task performance, and modulates neural plasticity through changes in glucoallostasis, In Control, Artificial

Intelligence, Robotics & Optimization (ICCAIRO), 2017. IEEE 2017; 325-9.

- 55. Lee RJ, Xiong G, Kofonow JM, et al. T2R38 taste receptor polymorphisms underlie susceptibility to upper respiratory infection. J Clin Invest 2012; 122:4145-59.
- 56. Sbarbati A, Tizzano M, Merigo F, et al. Acyl homoserine lactones induce early response in the airway. Anat Rec (Hoboken) 2009; 292:439-48.
- 57. Benford H, Bolborea M, Pollatzek E, et al. A sweet taste receptor-dependent mechanism of glucosensing in hypothalamic tanycytes. Glia 2017; 65:773-89.
- 58. Lazutkaite G, Soldà A, Lossow K, et al. Amino acid sensing in hypothalamic tanycytes via umami taste receptors. Mol Metab 2017; 6:1480-92.
- 59. Zhang Y, Hoon MA, Chandrashekar J, et al. Coding of sweet, bitter, and umami tastes: different receptor cells sharing similar signaling pathways. Cell 2003; 112:293-301.
- 60. Kojima I, Nakagawa Y. The role of the sweet taste receptor in enteroendocrine cells and pancreatic β -cells. Diabetes Metab J 2011; 35:451-7.
- 61. Ohkuri T, Yasumatsu K, Horio N, et al. Multiple sweet receptors and transduction pathways revealed in knockout mice by temperature dependence and gurmarin sensitivity. Am J Physiol Regul Integr Comp Physiol 2009; 296:R960-71.
- 62. Sprous D, Palmer RK. The T1R2/T1R3 sweet receptor and TRPM5 ion channel: Taste targets with therapeutic potential, In Progress in molecular biology and translational science. Academic Press 2010; 91:151-208.
- 63. Yasumatsu K, Ohkuri T, Sanematsu K, et al. Genetically-increased taste cell population with G-gustducin-coupled sweet receptors is associated with increase of gurmarin-sensitive taste nerve fibers in mice. BMC Neurosci 2009; 10:152.
- 64. Kojima I, Nakagawa Y, Ohtsu Y, et al. Sweet taste-sensing receptors expressed in pancreatic β-cells: sweet molecules act as biased agonists. Endocrinol Metab 2014; 29:12-9.
- 65. Alberini CM, Cruz E, Descalzi G, et al. Astrocyte glycogen and lactate: New insights into learning and memory mechanisms. Glia 2018; 66:1244-62.
- 66. Abraham WC, Dragunow M, Tate WP. The role of immediate early genes in the stabilization of long-term potentiation. Mol Neurobiol 1991; 5:297.
- 67. Sun X, Lin Y. Npas4: Linking neuronal activity to memory. Trends Neurosci 2016; 39:264-75.
- 68. Novak M, Halbout B, O'Connor EC, et al. Incentive learning underlying cocaine-seeking requires mGluR5 receptors located on dopamine D1 receptor-expressing neurons. J Neurosci 2010; 30:11973-82.
- 69. Gasbarri A, Pompili A, Clotilde Tavares M, et al.

Estrogen and cognitive functions. Expert Rev Endocrinol Metab 2009; 4:507-20.

- 70. McEwen B. Estrogen actions throughout the brain. Recent Prog Horm Res 2002; 57:357-84.
- Nyberg F, Hallberg M. Growth hormone and cognitive function. Nat Rev Endocrinol 2013; 9:357.
- 72. Lee AA, Owyang C. Sugars, sweet taste receptors, and brain responses. Nutrients 2017; 9:653.
- 73. Calvo SS, Egan JM. The endocrinology of taste receptors. Nat Rev Endocrinol 2015; 11:213.
- 74. Minatohara K, Akiyoshi M, Okuno H. Role of immediate-early genes in synaptic plasticity and neuronal ensembles underlying the memory trace. Front Mol Neurosci 2016; 8:78.
- 75. Goniotaki D, Lakkaraju AK, Shrivastava AN, et al. Inhibition of group-I metabotropic glutamate receptors protects against prion toxicity. PLoS Pathog 2017; 13:e1006733.
- 76. Niswender CM, Conn PJ. Metabotropic glutamate receptors: Physiology, pharmacology, and disease. Annu Rev Pharmacol Toxicol 2010; 50:295-322.
- 77. O'Keeffe J. Metabotropic glutamate receptors: Classification, structure and roles in disease. New York: Nova Science Publishers 2018.
- 78. Hanson JE, Smith Y. Group I metabotropic glutamate receptors at GABAergic synapses in monkeys. J Neurosci 1999; 19:6488-96.
- 79. Moriguchi S, Han F, Nakagawasai O, et al. Decreased calcium/calmodulin-dependent protein kinase II and protein kinase C activities mediate impairment of hippocampal long-term potentiation in the olfactory bulbectomized mice. J Neurochem 2006; 97:22-29.
- 80. Steinman MQ, Gao V, Alberini CM. The role of lactate-mediated metabolic coupling between astrocytes and neurons in long-term memory formation. Front Integr Neurosci 2016; 10:10.
- 81. Alberini CM. Transcription factors in long-term memory and synaptic plasticity. Physiol Rev 2009; 89:121-145.
- 82. Maddox SA, Monsey MS, Schafe GE. Early growth response gene 1 (Egr-1) is required for new and reactivated fear memories in the lateral amygdala. Learn Mem 2011; 18:24-38.
- 83. Tischmeyer W, Grimm R. Activation of immediate early genes and memory formation. Cell Mol Life Sci 1999; 55:564-74.
- 84. Hendrickx A, Pierrot N, Tasiaux B, et al. Epigenetic regulations of immediate early genes expression involved in memory formation by the amyloid precursor protein of Alzheimer disease. PLoS One 2014; 9:e99467.
- 85. Bramham CR, Alme MN, Bittins M, et al. The Arc of synaptic memory. Exp Brain Res 2010; 200:125-40.
- Korb E, Finkbeiner S. Arc in synaptic plasticity: From gene to behavior. Trends Neurosci 2011; 34:591-8.

- 87. Itoh N, Enomoto A, Nagai T, et al. Molecular mechanism linking BDNF/TrkB signaling with the NMDA receptor in memory: The role of Girdin in the CNS. Rev Neurosci 2016; 27:481-90.
- 88. Waltereit R, Dammermann B, Wulff P, et al. Arg3. 1/Arc mRNA induction by Ca2+ and cAMP requires protein kinase A and mitogen-activated protein kinase/extracellular regulated kinase activation. J Neurosci 2001; 21:5484-5493.
- 89. Nikolaienko O, Eriksen MS, Patil S, et al. Stimulusevoked ERK-dependent phosphorylation of activity-regulated cytoskeleton-associated protein (Arc) regulates its neuronal subcellular localization. Neuroscience 2017; 360:68-80.
- 90. Ying SW, Futter M, Rosenblum K, et al. Brainderived neurotrophic factor induces long-term potentiation in intact adult hippocampus: requirement for ERK activation coupled to CREB and upregulation of Arc synthesis. J Neurosci 2002; 22:1532-40.
- 91. Yao YL, Yang WM, Seto E. Regulation of transcription factor YY1 by acetylation and deacetylation. Mol Cell Biol 2001; 21:5979-5991.
- 92. Seto E, Yoshida M. Erasers of histone acetylation: The histone deacetylase enzymes. Cold Cold Spring Harb Perspect Biol 2014; 6:a018713.
- 93. Sultan FA, Day JJ. Epigenetic mechanisms in memory and synaptic function. Epigenomics 2011; 3:157-81.
- 94. Lubin FD, Gupta S, Parrish RR, et al. Epigenetic mechanisms: Critical contributors to long-term memory formation. Neuroscientist 2011; 17:616-32.
- 95. Singh P, Srivas S, Thakur MK. Epigenetic regulation of memory-Therapeutic potential for disorders. Curr Neuropharmacol 2017; 15:1208-21.
- 96. Day JJ, Sweatt JD. Epigenetic mechanisms in cognition. Neuron 2011; 70:813-29.
- 97. Zovkic IB, Guzman-Karlsson MC, Sweatt JD. Epigenetic regulation of memory formation and maintenance. Learn Mem 2013; 20:61-74.
- 98. Kitamura T, Sasaki T. Hypothalamic Sirt1 and regulation of food intake. Diabetol Int 2012; 3:109–12.
- 99. Cao Y, Yan Z, Zhou T, et al. SIRT1 regulates cognitive performance and ability of learning and memory in diabetic and nondiabetic models. J Diabetes Res 2017; 2017.
- 100. Welcome MO, Mastorakis NE, Pereverzev VA. Sweet-taste receptor signaling network and low-calorie sweeteners, In Sweeteners. Springer International Publishing 2016; 1-16.
- 101. Welcome MO, Pereverzev VA. A mini-review of the mechanisms of glucose memory enhancement. Int J Med Pharm Sci 2013; 4:17-30.
- 102. Dienel GA, Cruz NF. Aerobic glycolysis during brain activation: Adrenergic regulation and

influence of norepinephrine on astrocytic metabolism. J Neurochem 2016; 138:14–52.

- 103. Veldhuizen MG, Babbs RK, Patel B, et al. Integration of sweet taste and metabolism determines carbohydrate reward. Curr Biol 2017; 27:2476–85.
- 104. Boury-Jamot B, Halfon O, Magistretti PJ, et al. Lactate release from astrocytes to neurons contributes to cocaine memory formation. Bioessays 2016; 38:1266–73.
- 105. Boury-Jamot B, Carrard A, Martin JL, et al. Disrupting astrocyte-neuron lactate transfer persistently reduces conditioned responses to cocaine. Mol Psychiatry 2016; 21:1070–6.
- 106. Zhang Z, Gong N, Wang W, et al. Bell-shaped D-serine actions on hippocampal long-term depression and spatial memory retrieval. Cereb Cortex 2008; 18:2391–401.
- 107. Chan CB, Hashemi Z, Subhan FB. The impact of low and no-caloric sweeteners on glucose absorption, incretin secretion, and glucose tolerance. Appl Physiol Nutr Metab 2017; 42:793–801.
- 108. Murovets VO, Bachmanov AA, Zolotarev VA. Impaired glucose metabolism in mice lacking the Tas1r3 taste receptor gene. PLoS One 2015; 10:e0130997.
- 109. Gibbs ME. Role of glycogenolysis in memory

and learning: Regulation by noradrenaline, serotonin and ATP. Front Integr Neurosci 2015; 9:70.

- 110. Dienel GA. The metabolic trinity, glucoseglycogen-lactate, links astrocytes and neurons in brain energetics, signaling, memory, and gene expression. Neurosci Lett 2017; 637:18–25.
- 111. Pellerin L, Bouzier-Sore AK, Aubert A, et al. Activity-dependent regulation of energy metabolism by astrocytes: an update. Glia 2007; 55:1251–62.
- 112. Suzuki A, Stern SA, Bozdagi O, et al. Astrocyte-neuron lactate transport is required for long-term memory formation. Cell 2011; 144: 810–23.
- 113. Yang J, Ruchti E, Petit JM, et al. Lactate promotes plasticity gene expression by potentiating NMDA signaling in neurons. PNAS 2014; 111:12228–33.
- 114. Shah K, DeSilva S, Abbruscato T. The role of glucose transporters in brain disease: Diabetes and Alzheimer's disease. Int J Mol Sci 2012; 13:12629–55.
- 115. Dunn L, Allen GFG, Mamais A, et al. Dysregulation of glucose metabolism is an early event in sporadic Parkinson's disease. Neurobiol Aging 2014; 35:1111–5.