

Phytochemical Screening and Antibacterial Effect of *Stevia Rebaudiana (Bertoni*) Alcoholic Leaves Extract on *Streptococcus Oralis* (Dental Plaque's Primary Colonizer)

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ABSTRACT

Background: Medicinal herbs have drawn the attention of many researchers due to their anti-microbial and health promoting properties, especially after the increase in the side effects of chemical mouthwashes. "Stevia raebaudiana bertoni", a zero calorie sweetener native to South America, has been used by people for about 200 years to sweeten their drinks and treat some diseases. The anti-bacterial properties of Stevia extracts were studied by many researchers, but none of them investigated the effect of Stevia extract on Streptococcus oralis, a primary colonizer of dental plaque, which was addressed in this study.

Materials and methods: Supragingival plaque samples were cultured on MSA plate aerobically for 24 hours at 37°C. For identification and characterization of the isolated colonies, they were subjected to gram staining, hemolysis in Blood Agar Plate (BAP), catalase test, optochin sensitivity and Polymerase Chain Reaction (PCR) test. The alcoholic extract of Stevia leaves was prepared by maceration with 70% ethanol in a ratio of 1:10 (gm plant/ml solvent). Four experiments were involved in this study, the first one concerning the sensitivity of S. oralis to the study extract starting from 16 mg/ml to 512 mg/ml, with 0.2% CHX as a positive control and distilled water as a negative one. While the second experiment involved the determination of MBC, whereas the fourth experiment was done to determine some of the extract active ingredients by using High Performance Liquid Chromatography (HPLC) analysis.

Results: The diameter of inhibition zone for S. oralis increased with increasing the concentration of alcoholic extract of Stevia Rebaudiana bertoni and it is highest in chlorohexidine with significant difference (p<0.05). The MIC and MBC were 16 mg/ml and 32 mg/ml respectively. HPLC analysis revealed the presence of phytochemicals with antimicrobial activity including: Dihydroxy streptomycin: 0.86 ppm, rutin: 28.1 ppm, quercetin: 22.4 ppm and apigenin: 4.45 ppm.

Conclusions: The alcoholic extract of Stevia rebaudiana bertoni leaves had good antibacterial activity against S. oralis. 0.2% CHX demonstrated better antibacterial activity than the study extract, 512 mg/ml of Stevia extract had comparable effect to 0.2% CHX. HPLC analysis of Stevia rebaudiana bertoni alcoholic leaves extract revealed the presence of many active antibacterial constituents.

Key words: *Stevia rebaudiana bertoni,* Antibacterial effect, *S. oralis,* High Performance Liquid Chromatography (HPLC), Dihydroxy streptomycin

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INTRODUCTION

The human oral cavity is one of the most dynamic habitats for many bacterial species, where they compete fiercely to form multispecies biofilm structures. Dental plaque is an adherent bacterial film that is the primary pathological agent in periodontal diseases and dental caries. Pioneer microorganisms adhere to the pellicle, proliferate and form colonies, following that, secondary colonizers interact physiologically and physically, resulting in maturation of the dental plaque [1]. Inter bacterial binding to *streptococci* has major implications for the development of periodontal diseases. Many co adhering organisms are potential periodontal pathogens e.g. (*Porphyromons gingivalis, B. forsythus* and *Treponema denticola*). The fimbriae of *P. gingivalis* are able to bind to several streptococcal species, including *Streptococcus oralis, S. sanguinis, S. mitis* and *S. parasanguinis* [2]. Furthermore, it has been shown to localize on streptococcal rich plaque when introduced into the mouths of human volunteers [3]. On the other hand, later researches discovered that the gram positive 'early oral colonizers,' such as *Streptococcus* oralis, S. gordonii, S. sanguinis and Actinomyces naeslundii, co-aggregate with *F. nucleatum's* arginine inhabitable adhesion RadD [4].

There has been a constant increase in the search of natural anti-microbial agent which can be obtaining from green field instead of the chemical substances. Chlorhexidine is regarded as the "gold standard" antiplaque treatment and is particularly effective against gingivitis and widely used as an adjunct treatment for periodontitis but it unfortunately has several side effects including tooth and some restorations staining, unpleasant taste, sloughing of oral mucosa and supra gingival calculus formation enhancement [5].

The bioactive properties of medicinal plants have been target of an increasing interest. Stevia rebaudiana bertoni is a South American plant belongs to Asteraceae family; it is a natural sweetener nearly 300 times sweeter than sucrose and with the advantage of being non caloric and avoiding high blood sugar levels [6]. Many chemical constituents present in Stevia exert prominent biological effects, among them antioxidant, anti-diabetic (antihyperglycemic, insulin tropic and glucagonostatic), antimicrobial, antiplatelet, anti-cariogenic and antitumor properties [7,8]. de slavutzy conclude in his study that Stevia can act as anti-cariogenic and anti-gingivitis herb. It has the ability to inhibit the growth of certain bacteria and this explains the traditional use in the treatment of wounds, sores and gum diseases [9]. Furthermore, it has anti-inflammatory and anti-plaque effect [10].

High Performance Liquid Chromatography (HPLC) is a popular separative and analytical method which is ideal for separating a variety of bio chemicals, foods, heavy industrial chemicals and pharmaceuticals; it has many advantages over traditional methods such as: Speed of of quantification, reliability analysis, ease and automation. Because lower temperatures can be used and there are two competing phases (mobile and stationary) in HPLC versus one phase (the stationary phase) in Gas Chromatography (GC), HPLC can frequently achieve separations that GC cannot. Furthermore, there are many selective detectors available for use in HPLC, so a complete separation on the column is not required and a detector can be chosen to monitor only the species of interest [11].

Studies have been reported the antimicrobial effect of *Stevia* extracts on fungi and various bacteria. Hardly no studies have been reported the antibacterial activity of *Stevia rebaudiana bertoni* extract on *S. oralis*.

MATERIALS AND METHODS

Extraction of *Stevia rebaurdiana bertoni* leaves

Stevia leaves were obtained from and extracted in medical and aromatic plants research unit, college of agricultural engineering sciences, University of Baghdad. The alcoholic extract was prepared by maceration [12]. 100 gm powder of *Stevia* leaves were dissolved in 1000 ml of 70% ethanol concentration (conc.) with a ratio of 1:10 (gm plant/ml solvent). The filtered alcoholic extract

was spray dried to obtain alcoholic extract powder using laboratory mini spray dryer. The required concentrations were accordingly and sterile distilled water was used as a solvent.

S. oralis isolation and identification

The study was approved by the medical ethics committee, college of dentistry/University of Baghdad. Supra gingival plaque samples were taken from people (males and females) with a sterilized gracy curette; their approval was obtained prior to taking samples. Inclusion criteria included: Patients should not use antibiotics or mouth rinse within at least one month before the study. Each sample was immediately transferred to a sterile eppendrof tube containing 1.5 ml brain heart infusion broth [13].

For the identification and characterization of the isolated colonies, they were subjected to gram stain, hemolysis in Blood Agar Plate (BAP), catalase test, optochin sensitivity and Polymerase Chain Reaction (PCR) [14-18].

Polymerase Chain Reaction (PCR)

Genomic DNA was isolated from bacterial growth according to the protocol of "ABIO pure TM genomic DNA". Quantus fluorometer was used to detect the concentration of the extracted DNA. For 1 µl of DNA, 199 µl of diluted quantifluor dye was mixed. After 5 min incubation at room temperature, DNA concentration values were detected. Primers were supplied by macrogen company in a lyophilized form, *S. oralis* forward: 5`-TCCCGGTCAGCAAACTCCAGCC 3`, *S. oralis* reward: 5`-GCAACCTTTGGATTTGCAAC-3`. Annealing temperature: 66°C, Product size (Pb): 374.

Lyophilized primers were dissolved in 300 µl of nuclease free water to give a final concentration of 100 pmol/ μ l as a stock solution. A working solution of the primer was prepared by adding 10 µl of primer stock solution (stored at freezer -20°C) to 90 µl of nuclease free water to obtain a working primer solution of 10 pmol/µl. For each sample, the following component were calculated according to manufacturer's instructions and mixed in a tube to achieve a total volume of 20 µl: 10 µl master mix, 1 µl of 10 pmol/µl F primer, 1 µl of 10 pmol/µl R primer, 6 μ l of nuclease free water and 2 μ l of 22 ng/ μ l of sample DNA. According to manufacturer's instructions, the calculated tubes of PCR components were applied in the PCR device for the amplification of the accused DNA sample, the reaction volume/run was 20, number of PCR cycles were 30. After DNA amplification, agarose gel electrophoresis was adopted to confirm the presence of S. oralis DNA. The ethidium bromide stained bands in gel were visualized using gel imaging system.

Prepartion of *S. oralis* suspension was done by direct colony suspension method according to cavalieri follows by an inoculating loop, isolated colonies were picked up from the agar plate (not older than 18-24 hours) and suspended in 1 ml Mueller Hinton broth (MHA) then vortexes and adjusted by absorbance micro plate reader to an absorbance of 0.600 at 600 nm which is

corresponds to 0.5 McFarland standard (equivalent to 1 X 10^8 cells/ml) [19].

Testing the sensitivity of *Streptococcus oralis* to alcoholic *Stevia rebaudiana bertoni* leaves extract

Agar well diffusion method was applied to study the antibacterial effects of different concentrations of alcoholic *Stevia* extract (16 mg/ml, 32 mg/ml, 64 mg/ml, 128 mg/ml, 256 mg/ml and 512 mg/ml) on Mueller Hinton agar.

This test was done according to cavalieri follows two wells of equal depth and size (6 mm in diameter) were made in each agar plate using Pasteur pipette under aseptic conditions. Each well filled with 50 μ l of each concentration. Non-alcoholic 0.2% chlorhexidine gluconate was used as positive control and sterile distilled water was the negative one. The plates were incubated aerobically at 37°C for 24 hours. Using a ruler, the diameter of the inhibition zone across each well was measured in millimeters.

Minimum Inhibitory Concentration (MIC) and Minimum Bactericidal Concentration (MBC)

The MIC experiment was done by two fold serial broth micro dilution. 100 μ l of BHI broth was added into each nine wells of two rows, labeled W1 (well 1) to W9, in a 96 well micro titer plate. 100 µl of stevia extract with a concentration of 512 mg/ml was added into each W1 of the respective row and then two fold serial dilutions was done from W1 through W9. W10 of the first row contained 100 µl broth and 100 µl of 0.2% CHX while W11 and W12 contained 100 µl Mueller Hinton Broth (MHB) only, the entire first row was considered to be a blank row (without bacteria). All wells of the second row were inoculated with 100 μ l of bacterial suspension. W10 of the second row was contained 100 μ l of 0.2% CHX in broth and 100 μ l bacterial suspension and it was considered the positive control, while W11, W12 was contained 100 μI MHB and 100 μI bacterial suspension and these wells were considered to be the negative control. The micro titer plate covered and incubated aerobically overnight at 37°C. Following incubation, the wells were examined for turbidity and checked by an "absorbance micro plate reader" adjusted to (600 nm) wave length.

For the determination of MBC, an aliquots of $50 \,\mu$ l from the MIC well and from the well proceeded it were seeded on Brain Heart Agar (BHA) plates and incubated for 24 h at 37°C. Then the agar plates were observed for the presence or absence of bacteria [20].

High Performance Liquid Chromatography (HPLC) test for identifying and quantifying some of the active constituents in alcoholic *Stevia rebaudiana* leaves extract

This test was done in ministry of science and technology, department of material research according to Gini and Jeya Jothi with slight modification [21].

Stevia alcoholic extract was prepared for HPLC test to detect and quantitate each of: Streptomycin, rutin, quercetin and apigenin. The standard concentration for all of them was 5 ppm except for streptomycin it was 1 ppm. 1 gm of the sample (Stevia rebaudiana alcoholic extract) dissolved in 5 ml acetonitrile (HPLC grade) in a glass jar, mixed and closed by paraffin wax, the jar vortexed and putted in ultrasonic path at 35°C for 15 min. The solution was filtered by 0.45 micro filter, 20 μ l of the sample was injected in HPLC binary pump device at the following conditions: Mobile phases: A-acetonitrile: 0.5% formic acid=(70:30 v/v). B-acetonitrile: 0.5% formic acid=(30:70 v/v). Time (min): 0.01, 4, 8, 12, 15, 20. Gradient program (conc B): 80%, 80%, 90%, 90%, 80% stop. Flow rate: 0.5 ml/min. Detector: UV-VIS at 280 nm. Column stationary phase: ODS-C₁₈ (150 x 4.6 I.D) mm, 5 µm particle size. Column temperature: 25°C.

The concentration of the sample was calculated by the following formula:

"Conc. of sample =
$$\frac{conc.of \ standard \times Area \ of \ sample}{Area \ of \ standard} \times dilution \ factor"$$

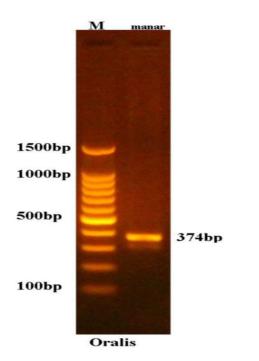
Statistical analysis

Data description, analysis and presentation were performed using minimum, maximum, mean, Standard Deviation (SD), Standard Error (SE), Shapiro Wilk test, Levene test and One Way Analysis of Variance (ANOVA) by using Statistical Package for social Science (SPSS version 22, Chicago, Illionis, USA). Not significant P>0.05, significant P<0.05.

RESULTS

Identification of Streptococci

Colonies of *S. oralis* appeared spherical in shape with shiny dark blue color, smooth raised convex surface and easily removed from the agar surface. Biochemical results confirmed that the study bacterium is a gram positive, alpha hemolytic, optochin resistant and catalase negative *streptococci.* PCR result confirmed the identification of *S. oralis* (Figure 1).



Results of the experiments

Findings from the statistical analysis show that the diameter of inhibition zone for *S. oralis* increased with increasing the concentration of alcoholic extract of *Stevia rebaudiana bertoni* and it is highest in chlorohexidine with significant difference (p<0.05), further more using multiple comparisons between groups by Tukey's HSD each group when compared against other group and with chlorohexidine, the result showed significance difference (p<0.05). The results are shown in Tables 1-3. The MIC and MBC of the alcoholic *Stevia rebaudiana bertoni* extract against *S. oralis* were 16 mg/ml and 32 mg/ml respectively, 0.2% CHX (0.1% in broth) revealed bacteriostatic effect.

Figure 1: Result of the amplification of *S. oralis* DNA. M: 100 bp ladder marker; Bp: Base pair.

Groups (mg/ml)	Mean	± SD	± SE	Minimum	Maximum
16	11.3	0.447	0.2	11	12
32	13.7	0.447	0.2	13	14
64	16.3	0.447	0.2	16	17
128	18.5	0.5	0.224	18	19
256	20.9	0.652	0.292	20	21.5
512	24.6	0.418	0.187	24	25
СНХ	26.7	0.447	0.2	26	27
		Levene test=0.437	, p value=0.848 NS		

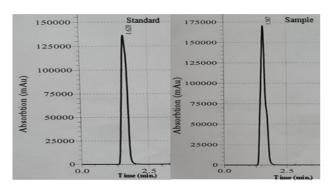
Table 2: Statistical test of diameter of inhibition zone for *S. oralis* among groups using One Way Analysis of Variance (ANOVA).

ANOVA							
	Sum of squares	Df	Mean square	F	P value		
Between groups	945.186	6	157.531	668.313	0.000 Sig		
Within groups	6.6	28	0.236				
Total	951.786	34					
		Sig=Signif	ïcant at p<0.05.				

Groups (n	ng/mL)	Mean difference	P value	
16	32	-2.4	0.00000	
_	64	-5	0.00000	
_	128	-7.2	0.00000	Sig
_	256	-9.6	0.00000	
_	512	-13.3	0.00000	
_	СНХ	-15.4	0.00000	
32	64	-2.6	0.00000	
_	128	-4.8	0.00000	
_	256	-7.2	0.00000	
_	512	-10.9	0.00000	
_	СНХ	-13	0.00000	
64	128	-2.2	0.00000	
_	256	-4.6	0.00000	
_	512	-8.3	0.00000	
_	СНХ	-10.4	0.00000	
128	256	-2.4	0.00000	
_	512	-6.1	0.00000	
_	СНХ	-8.2	0.00000	
256	512	-3.7	0.00000	
_	СНХ	-5.8	0.00000	
512	СНХ	-2.1	0.00000	

Table 3: Multiple comparisons of diameter of inhibition zone for S. Oralis between groups usingtukey HSD.

High performance liquid chromatography revealed the presence of active antimicrobial phytochemicals with the following concentrations: Dihydroxy streptomycin 0.86 ppm (Figure 2); rutin 28.1 ppm, quercetin 22.4 ppm and



apigenin 4.45 ppm (Figure 3).

Figure 2: HPLC revealed the presence of dihydroxy streptomycin in *Stevia rebaudiana bertoni* alcoholic leaves extract.

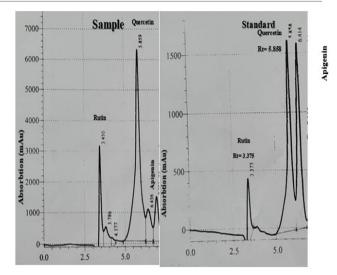


Figure 3: HPLC of alcoholic *Stevia rebaudiana bertoni* leaves extract revealed the presence of Rutin, Querctin and Apigenin. RT stands for retention time.

DISCUSSION

Medicinal herbs have been used for centuries to treat human diseases due to their health promoting properties. It has been estimated that 80% of the population still use them as complementary and alternative medicines because of its safety and effectively, as a result, scientists began to look into the biological activities of medicinal plants especially after the emergence of microbial resistance and side effects of the chemical drugs [22,23].

Stevia rebaudiana bertoni has many nutritional and therapeutic properties and it is considered a potential drug candidates. Antimicrobial activity screening studies of Stevia extracts showed enough activity to inhibit the growth of pathogenic bacteria like E. coli, Salmonella typhi, Enterococcus faecalis, Bacillus subtilis, Pseudomonas Proteus mirabilis, Vibrio aeruginosa, cholerae, Staphylococcus Aeromonas hvdrophila. aureus, Streptococcus mutuns and Lactobacillus. Also it has anticariogenic, anti-gingivitis, anti-plaque, anti-inflammatory and healing properties [24]. For the previously mentioned properties and because there is no study on the antibacterial activity of Stevia on S. oralis in comparison to 0.2% CHX, the present study was conducted and may be considered as the first report.

The present study found a concentration dependent antibacterial activity of the alcoholic *Stevia* extract against *S. oralis* in that the diameter of inhibition zone increases as the concentration of the extract increases, this could be attributed to the increase in the amount of the dissolved active ingredients of the extract.

The alcoholic *Stevia* extract showed promising results; the mean value of Inhibition zone of the alcoholic *Stevia* extract at 512 mg/ml against *S. oralis* (24.6 mm) was somewhat comparable to that of 0.2% CHX against *S. oralis* (26.7 mm), it could be considered a satisfactory result when we weight the benefits of *Stevia* against the side effects of CHX. The MIC and MBC of alcoholic *Stevia* extract against the study bacteria were found to be 16 mg/ml and 32 mg/ml respectively, these results indicate that *Stevia* extract exert bactericidal action according to levison as mentioned in mogana in which stated that the agent exhibit bactericidal action if the mbc/mic ratio \leq 4 otherwise it is considered a bacteriostatic drug [25].

The precise antimicrobial action of *Stevia* is unknown, but it could be linked to the presence of many antimicrobial components such as: Diterpene glycosides, flavonoids, tannins, saponins, dihydrodeoxy streptomycin and polyphenols [26-30]. And the synergism between the phytochemical compounds in the extract. It is more difficult to develop bacterial resistance through genetic mutations triggered by external stimuli because of the association of numerous different molecules in plant extracts [31].

High performance liquid chromatography analysis of alcoholic *Stevia* extract revealed the presence of many active constituents including: Dihydroxy streptomycin, apigenin, quercetin and rutin. Dihydroxy streptomycin found with a concentration of (0.86 ppm) in the study extract. It exerts its antibacterial action by inhibiting the bacterial proteins biosynthesis [32]. While rutin occur at a relatively high concentration (28.1 ppm), which exhibits a good antibacterial property, it is thought to alter glucose and amino acid metabolism, protein synthesis and cell wall integrity [33]. It has been reported that rutin exhibits antibacterial action against *Staphylococcus aureus, Eschericia coli* and *Staphylococcus glurance* [34]. Also against *Klebsiella pneumonia* and *Pseudomonas aeruginosa* [35].

Quercetin is a flavonoid with anti-bacterial properties, it is found in the study extract at a concentration of (22.4 ppm). According to Siriwong quercetin has a synergistic impact with amoxicillin against Amoxicillin Resistant Staphylococcus Epidermidis (ARSE) through four mechanisms: Inhibiting peptidoglycan production and βlactamases activity, increasing cell membrane permeability and protein amide I and II and decreasing fatty acids in bacterial cells. It also demonstrated strong antibacterial efficacy against a broad spectrum pathogens responsible for hospital and community acquired infections by inhibiting bacterial DNA gyrase and topoisomerase IV [37].

Apigenin is a flavone occurs naturally in the medicinal plants, it is found in the study extract at a concentration of (4.45 ppm). Kim implies that apigenin causes bacterial death *via* activating cellular oxidative pathways that are reliant on Reactive Nitrogen Species (RNS) and Reactive Oxygen Species (ROS) generation and accumulation [38].

CONCLUSION

The ethanolic *Stevia rebaudiana bertoni* leaves extract has good antibacterial effect against *S. oralis* starting from 16 mg/ml to 512 mg/ml but 0.2% CHX demonstrated better antibacterial activity. HPLC analysis revealed the presence of many active antibacterial constituents. Further studies concerning its antimicrobial effect on periodontal pathogens are required to validate its usage as supportive oral health measure.

CONFLICTS OF THE INTEREST

No conflicts of the interest

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