

# The Alterations in Some Salivary Biomarkers and Oral Microbiome in COVID-19 Patients and Healthy Individuals

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## ABSTRACT

A saliva samples have been collected from 42 COVID-19 patient and a 42 healthy individuals and tested for salivary biomarkers of,  $\alpha$ -amylase, lysozyme in addition to and mutans streptococci. Results showed non-significant difference between mean concentration of  $\alpha$ -amylase in COVID-19 patients (5.48  $\mu$ /ml) and healthy individuals (6.05  $\mu$ g/ml), high significant difference in mean concentration of lysozyme in COVID-19 patient (170.42  $\mu$ g/ml) and healthy individuals (8.47  $\mu$ g/ml), and non-significant difference in mean concentration of mutans streptococci in COVID-19 patients (6.88)  $\times 10^6$  CFU/ml and healthy individuals (6.58  $\times 10^6$  CFU/ml).

**Key words:** COVID-19, SARS-CoV-2, Salivary lysozyme, Salivary  $\alpha$ -amylase, *Mutans streptococci*

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## INTRODUCTION

Saliva is a bio-fluid constituted of various fluids of secretions of salivary glands, respiratory secretions, crevicular and exfoliated epithelial cells. Presence of SARS-CoV-2 in saliva is attributed to the viral replication and RNA secretion in any cells and tissues related to the production of salivary components like salivary glands, periodontal tissues and the upper respiratory tract cells [1].

The microbial constituents of a eubiotic Human Oral Microbiome (HOM) can inhibit colonization of pathogens through competitive exclusion and/or via facilitation of immune response to exclude the pathogen [2,3]. It is reported that a vital collaborative interactions happen between viruses and the micro biome and that the micro biome could be regulate and it, in turn, could be regulated by viruses through various mechanisms [4]. The oral micro biota can produce anti-viral substances (defensins) against various viruses, including respiratory tract viruses like coronaviruses, herpes viruses, adenoviruses, orthomyxoviruses and papillomaviruses [5]. Otherwise, invading viruses could results in dysbiosis and progression of disease [6].

The pandemic of coronavirus SARS-CoV-2, the causative of COVID-19 is respiratory virus that invades the oropharynx

as the primary site of replication but the possible impact on oral micro biome through development of infection remains un-clarified. In particular, there are no date available concerning the non-bacterial constituents of HOM (fungi and viruses), that have been shown vital for other diseases. Regarding COVID-19, it is reported that the presence of gingival inflammation/periodontitis was associated with a 3.5-fold increase in the risk for admission to Intensive Care Units (ICU), a 4.5-fold increase in the probability for assisted ventilation and a risk of 8.81-fold increase in the probability for death as a consequences of COVID-19, separately from any other concomitant risk factors [7].

The functions of salivary biomarker of  $\alpha$ -amylase extensively studied for their biological activities but their correlation to each other in COVID-19 patient and healthy individuals to COVID-19 are still unrevealed. The current study is designed to assess whether there is any association between salivary  $\alpha$ -amylase, lysozyme, and total a count of *mutans Streptococci*. The study aimed to measure the salivary  $\alpha$ -amylase level in both groups (COVID-19 patients and healthy individuals), to measure the salivary lysozyme level in both groups (COVID-19 patients and control group), to calculate the total viable count of *mutans Streptococci* and *C. albicans* among COVID-19 patients and to evaluate the relationship between salivary  $\alpha$ -amylase, lysozyme, and total viable a count of *mutans Streptococci* among COVID-19 patients.

## MATERIALS AND METHODS

**Saliva collection:** After taking patients agreement for collecting saliva samples, checking that the patients didn't

take antibiotics or any medications for the latest two weeks, then giving the patients plain tubes that numbered before sample collection. Saliva sample collection was made in early morning at time between 8 am to 10 am.

The amount of the collected saliva was between 1-3 ml of un-stimulated saliva by allowing the saliva to accumulate in the mouth and then spitting into a tube. After collecting the salivary sample from each patient, the tubes were placed in a cool box with ice to transfer them to the laboratory to be cultured within less than an hour, then 0.1 ml would be taken from the salivary sample by micro pipette for the serial dilution tubes using PBS. The resting saliva was centrifuged for 3000 rpm for 10 min, and the clear supernatant was e and stored in freezer at -20°C until the determinations of salivary  $\alpha$ -Amylase, lysozyme and melatonin were done by ELISA test

### Determination of lysozyme level

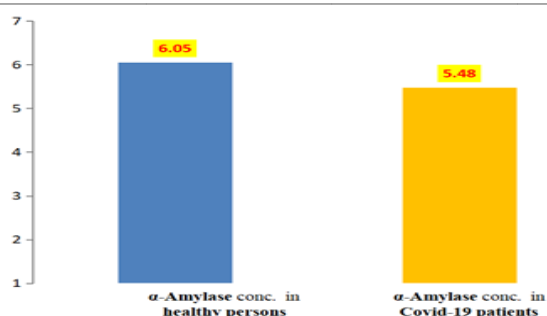
This kit was based on Competitive-ELISA detection method (Cell Biolabs, USA). The microtiter plate provided in this kit has been pre-coated with antibody. During the reaction, target in the sample or standard competes with a fixed amount of Biotin-Antigen. Excess conjugate and unbound sample or standard are washed from the plate. HRP-Streptavidin was added and unbound conjugates were washed away with wash buffer. Then TMB substrate solution is added to each well. The enzyme substrate reaction is terminated by the addition of a sulfuric acid solution and the colour change is measured spectrophotometrically at a wavelength of 450 nm. The concentration of target in the samples is then determined by comparing the OD of the samples to the standard curve.

### Determination of salivary $\alpha$ -Amylase

The ELISA test kit (LDN, Germany) provides a quantitative *in vitro* assay for free  $\alpha$ -amylase in human saliva. The test kit contains microtiter strips each with 8 break off reagent wells coated with anti-rabbit antibodies. In the first reaction step, diluted patient samples are pipetted into the reagent wells together with peroxidase labelled  $\alpha$ -amylase and a specific rabbit anti- $\alpha$ -amylase antibody.  $\alpha$ -amylase from the patient sample

**Table 1: Descriptive of  $\alpha$ -amylase.**

Variable	No. of cases	Minimum	Maximum	Mean	SE	SD
Healthy individuals	42	3.6	9.09	6.05	0.25	1.66
COVID-19 patients	42	1.79	11.2	5.48	0.33	2.18



**Figure 1: Means of  $\alpha$ -amylase among COVID-19 patients and healthy individuals groups.**

and the labelled  $\alpha$ -amylase in the conjugate compete for the free binding sites of the specific antibody. In the third incubation step, the bound peroxidase catalyses a colour reaction with the peroxidase substrate Tetra Methyl Benzidine (TMB). The intensity of the colour formed is inversely proportional to the concentration of  $\alpha$ -amylase in the sample. The results for the samples are determined using the standard curve.

### Cultivation of *mutans streptococci*

Saliva samples were centrifuged at 3000 rpm for 10 min. Precipitate has been discarded and the supernatant forwarded for culture and identification of *mutans Streptococci* and The supernatant has been serially 10-folds diluted in PBS enumeration of *mutans streptococci*, the supernatant of saliva was serially 10-folds diluted in PBS and streaked on MSBA agar for calculation of CFU/ml of *Mutans Streptococci*.

**Statistical analysis:** Statistical analysis and processing of the data were carried out using SPSS version 21 (Statistical Package for Social Sciences) under Windows 10. Data were subjected to the following:

### Descriptive statistics

- Arithmetic Means (M), Standard Deviation (SD) and Standard Error (SE).
- Minimum (mini) and maximum (maxi).

### Inferential statistics

The statistical tests that were used in this study:

- Anova test.
- L.S.D. test.
- Student's t-test and.
- Pearson correlation coefficients.

The level of significance was accepted at  $P < 0.05$ , and highly significance when  $P < 0.01$ .

## RESULTS

**$\alpha$ -Amylase:** Salivary  $\alpha$ -amylase level in COVID-19 patients group and healthy individuals group is shown in Table 1 and Figure 1, which revealed the presence of mean value of salivary  $\alpha$ -amylase among the COVID-19 patients group (5.48) u/ml less than healthy individuals group (6.05) u/ml, with a statistical non-significant difference between the two groups, the t-value was (1.602) and the p value was (0.117).

**Lysozyme:** Salivary lysozyme level in COVID-19 patients group and healthy individuals group is shown in Table 2 and Figure 2, which revealed the presence of higher mean value of salivary lysozyme among the COVID-19 patients group (170.42) g/ml than healthy individuals group (8.47) g/ml, with a statistical high significant difference between the two groups, the t-value was (7.668) and the p value was (0.001).

Table 2: Descriptive of lysozyme.

Variables	No. of cases	Minimum	Maximum	Mean	SE	SD
COVID-19 patients	42	24.38	513.3	170.42	21.11	136.86
Healthy individuals	42	4.76	9.29	8.47	0.16	1.07

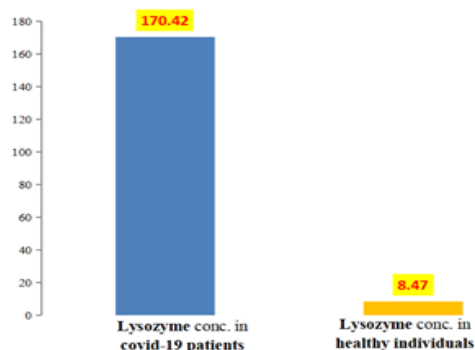


Figure 2: Means of lysozyme among COVID-19 patients and healthy individuals group.

Table 3: Descriptive of mutans streptococcus.

Variables	No. of cases	Minimum	Maximum	Mean	SE	SD
Healthy individuals	42	2.8	11.28	6.58	0.36	2.39
COVID-19 patients	42	3	17.44	6.88	0.55	3.61

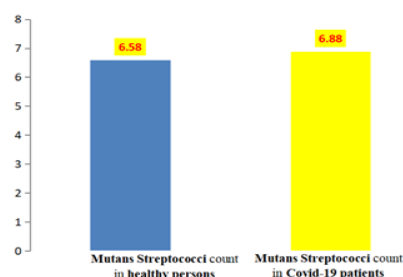


Figure 3: Means of Total viable count of salivary mutans streptococci among COVID-19 patients and healthy individuals group.

## DISCUSSION

### Salivary lysozyme level in COVID-19 patients group and healthy individuals group

The presence of lysozyme in body fluids, including saliva, is an important factor in non-specific mechanisms towards microbial infections [19-21]. The high significant difference between the COVID-19 patients and healthy individuals groups is a strong marker that SARS-CoV-2 provokes the salivary glands to secrete a 20 fold of lysozyme concentration of healthy individuals to counteract the viral infection.

Lysozyme is bactericidal for gram-positive bacteria via hydrolyzing the  $\beta$ -1,4 glycosidic bond between N-acetylglucosamine and N-acetylmuramic acid of the bacterial cell wall [22]. Furthermore, the lysozyme exerts antimicrobial activity through binding to negatively charged surfaces of microbes owing to its cationic nature [23,24]. The immunomodulatory action of lysozyme has only been appreciated in last few years.

Within neutrophils and macrophages, lysozyme works to increase their pro-inflammatory response, but when it secreted outside the above mentioned cells as well as epithelial cells, it limits inflammation through decreasing the chemotaxis and oxidative burst in neutrophils [25], it suppresses the production of IL-6 and TNF- $\alpha$  in

### Total Viable Count of Mutans Streptococci (CFU/ml)

Result in Table 3 and Figure 3 shows the statistical analysis of viable count (CFU/ml)  $\times 10^6$  of Total Viable Count of Salivary *Mutans streptococcus*, there was Non-significant difference between two groups, the t-value was (0.43), the p value (0.66), the mean value in healthy individuals group (6.58)  $\times 10^6$  CFU/ml was less than COVID-19 patients group mean value (6.88)  $\times 10^6$  CFU/ml

macrophages [26], it binds and decreases the circulating levels of Advanced Glycation End products (AGEs) (that are pro-oxidative) in addition to increasing their renal excretion [27] and it disrupts the capacity of peptidoglycan to bind the complement factors that works as anaphylotoxins [22]. Moreover, when subjected to artificial conditions of gastro-intestinal conditions, the hydrolyzate of lysozyme of hen egg white showed remarkable Angiotensin Converting Enzyme (ACE) and anti-oxidant activity [28,29]. As mentioned above, the oxidative stress (including participation of AGEs), cytokines of IL-6 and TNF- $\alpha$ , inflammation caused by macrophages and neutrophils and the Renin-Angiotensin System (RAS) are characteristic in the Acute Respiratory Distress Syndrome (ARDS) and/or severe COVID-19. It is most interested that the activity of lysozyme beside lactoferrin in neuroprotection of Alzheimer's patients through inhibition of amyloid-beta aggregation [30] could be an active mechanism in treatment of potential neurological manifestations in severe cases of COVID-19.

The increase in Salivary Antimicrobial Proteins (sAMPs) like lysozyme and lactoferrin in Lower Respiratory Tract Infection (LRTI) was reported in several previous studies [31,32]. The advantage of increasing level of lysozyme is to raise the immunomodulatory effects of lysozyme to counteract SARS-CoV-2 infection [25-28]

### The viable total count of salivary mutans streptococci in COVID-19 patients group and healthy individuals group

The count concentration of *mutans streptococci* in COVID-19 patients and healthy individuals was attributed to the fact that the natural habitat of *mutans streptococci* is the oral cavity. The *mutans streptococci* have the ability to exert effective persistence in oral cavity due to formation of diverse human associated-biofilms [33-65] reported that the bacterial profile of oral microbiota in COVID-19 patients is characterized by dominance of *Prevotella salivae* and *Veillonella infantium*, whereas *Neisseria perflava* and *Rothia mucilaginosa* were dominant in healthy individuals along with *N. perflava*, *K. gabonensis*, *G. elegans*, *Porphyromonas pasteri*, *Gemella taiwanensis*, *R. mucilaginosa*, and *Streptococcus oralis*. *Mutans streptococci* concentration in oral cavity is unaffected by COVID-19 [66-95].

### CONCLUSION

It is concluded that salivary lysozyme is highly responsive through COVID-19 to immunomodulate the infection. There is an inverse relationship between salivary melatonin level and COVID-19 that is resulted, probably, from fast utilization of melatonin as free radical scavenger to counteract the inflammation-generated high level of ROS. The secretory function of salivary glands for secretion of  $\alpha$ -amylase is unaffected during COVID-19 due to the homeostasis of the enzyme in both COVID-19 patients and healthy individuals. Oral candidiasis is a sequelae of COVID-19 as result of saliva low flow rate (Xerostomia) occurred during SARS-CoV-2 infection to salivary glands. The total count of *Mutans Streptococci* is stable during COVID-19.

### RECOMMENDATIONS

It is recommended to study the effect of COVID-19 on oral microbiota of gram-positive and gram-negative bacteria and monitor the potential presence of bacteremia and/or septicemia in COVID-19 patients due to overgrowth of any member of oral bacteria, study level of other salivary biomarkers through course of COVID-19 pathology like C-Reactive Protein (CRP), myoglobin, creatine kinase isoform MB,  $\alpha$ -2-macroglobulin, glycosylated hemoglobin (HbA1c) and various Interleukins (ILs), study the involvement of periodontal diseases in the COVID-19 due to depletion of salivary melatonin level in COVID-19 patients that is correlated to periodontal diseases, study the correlation between severity of COVID-19 and levels of salivary biomarkers and oral micro biome to uncover the contribution of salivary biomarkers and oral micro biome in determination of susceptibility of an individual to COVID-19.

### CONFLICT OF INTEREST

The author declare that there are no conflicts of interest

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### REFERENCES

- Chen T, Hudnall SD. Anatomical mapping of human herpesvirus reservoirs of infection. *Mod Pathol* 2006; 19:726-737.
- Wilks J, Beilinson H, Golovkina TV. Dual role of commensal bacteria in viral infections. *Immunol Rev* 2013; 255:222-229.
- Wilks J, Golovkina T. Influence of microbiota on viral infections. *PLoS Pathog* 2012; 8:1002681.
- Li N, Ma WT, Pang M, et al. The Commensal Microbiota and Viral Infection: A Comprehensive Review. *Front Immunol* 2019; 10:1551.
- Pfeiffer JK, Sonnenburg JL. The intestinal microbiota and viral susceptibility. *Front Microbiol* 2011; 2:92.
- Lynch SV. Viruses and microbiome alterations. *Ann Am Thorac Soc* 2014; 11:57-60.
- Marouf N, Cai W, Said KN, et al. Association between periodontitis and severity of COVID-19 infection: A case-control study. *J clin periodontal* 2021; 48:483-491.
- Granger DA, Kivlighan KT, El-Sheikh M, et al. Salivary alpha-amylase in biobehavioral research: Recent developments and applications. *Ann N Y Acad Sci* 2007; 1098:122-144.
- Chrousos GP, Gold PW. The concepts of stress and stress system disorders: Overview of physical and behavioral homeostasis. *Erratum in JAMA* 1992; 267:244-1252.
- Kirschbaum C, Read GF, Hellhammer DH. Assessment of hormones and drugs in saliva in biobehavioral research. *Gottingen: Hogrefe and Huber* 1994.
- Scannapieco FA, Torres G, Levine MJ. Salivary  $\alpha$ -amylase: Role in dental plaque and caries formation. *Crit Rev Oral Biol Med* 1993; 4:301-307.
- Rogers JD, Palmer RJ Jr, Kolenbrander PE. Role of *Streptococcus gordonii* amylase-binding protein A in adhesion to hydroxyapatite, starch metabolism, and biofilm formation. *Infect Immun* 2001; 69:7046-7056.
- Matuck BF, Dolnikoff M, Duarte-Neto AN, et al. Salivary glands are a target for SARS-CoV-2: a source for saliva contamination. *J Pathol* 2021; 254:239-243.
- Chen M, Shen W, Rowan NR, et al. Elevated ACE2 expression in the olfactory neuroepithelium: implications for anosmia and upper respiratory SARS-CoV-2 entry and replication. *bioRxiv : the preprint server for biology* 2020.
- Song G, Liang G, Liu W. Fungal Co-infections Associated with Global COVID-19 Pandemic: A Clinical and Diagnostic Perspective from China. *Mycopathologia* 2020; 185:599-606.
- Song J, Li Y, Huang X, et al. Systematic analysis of ACE2 and TMPRSS2 expression in salivary glands reveals underlying transmission mechanism caused by SARS-CoV-2. *J Med Virol* 2020; 92:2556-2566.
- Farshidfar N, Hamedani S. Hyposalivation as a potential risk for SARS-CoV-2 infection: Inhibitory role of saliva. *Oral diseases* 2021; 3:750-751.
- Pribadi RR, Simadibrata M. Increased serum amylase and/or lipase in coronavirus disease 2019 (COVID-19) patients: Is it really pancreatic injury. *JGH Open* 2021; 5:190-192.
- Jenzano JW, Hogan SL, Lundblad RL. Factors influencing measurement of human salivary lysozyme in lysoplate and turbidimetric assays. *J clin microbiol* 1986; 24:963-967.
- Nasiri K. COVID-19 and the Antiviral Effect of Saliva. *Eur J Dent* 2020; 14:177-178.

21. Pedrosa MS, Sipert CR, Nogueira FN. Salivary Glands, Saliva and Oral Findings in COVID-19 Infection. *Odontopediatria Clín. Integr* 2020; 20:0104.
22. Ragland SA, Criss AK. From bacterial killing to immune modulation: Recent insights into the functions of lysozyme. *PLoS Pathog* 2017; 13:1006512
23. Sava G. Pharmacological aspects and therapeutic applications of lysozymes. *Lysozymes: Model Enzymes in Biochemistry and Biology*. Jolles P. (Lysozyme: Model Enzymes in Biochemistry and Biology). Birkhäuser Verlag, Basel, Switzerland, 1996; 75:433-449.
24. Ibrahim HR, Imazato K, Ono H. Human lysozyme possesses novel antimicrobial peptides within its N-terminal domain that target bacterial respiration. *J Agric Food Chem* 2011; 59:10336-10345.
25. Gordon LI, Douglas SD, Kay NE, et al. Modulation of neutrophil function by lysozyme. Potential negative feedback system of inflammation. *J Clin Invest* 1979; 64:226-232.
26. Tagashira A, Nishi K, Matsumoto S, et al. Anti-inflammatory effect of lysozyme from hen egg white on mouse peritoneal macrophages. *Cytotechnol* 2018; 70: 929-938.
27. Aldini G, Vistoli G, Stefek M, et al. Molecular strategies to prevent, inhibit, and degrade advanced glycoxidation and advanced lipoxidation end products. *Free radical res* 2013; 47:93-137.
28. Rao S, Sun J, Liu Y, et al. ACE inhibitory peptides and antioxidant peptides derived from *in vitro* digestion hydrolysate of hen egg white lysozyme. *Food Chem* 2012; 135:1245-1252.
29. Kaabi SAG. Prophylactic Phage Therapy in Infant Rabbits Model of cholera. *Karbala Int J Mod Sci* 2021; 7:2.
30. Helmfors L, Boman A, Civitelli L, et al. Protective properties of lysozyme on  $\beta$ -amyloid pathology: implications for Alzheimer disease. *Neuro biol Dis* 2015; 83:122-133.
31. Akinbi HT, Epaud R, Bhatt H, et al. Bacterial killing is enhanced by expression of lysozyme in the lungs of transgenic mice. *J immunol* 2000; 165:5760-5766.
32. Canto E, Roca E, Perea L, et al. Salivary immunity and lower respiratory tract infections in non-elite marathon runners. *PloS one* 2018; 13.
33. Kreth J, Merritt J, Qi F. Bacterial and host interactions of oral streptococci. *DNA and cell biology* 2009; 28:397-403.
34. Iebba V, Zanotta N, Campisciano G, et al. Profiling of Oral Microbiota and Cytokines in COVID-19 Patients. *Front microbiol* 2021; 12:1603.
35. J Zeng L, Kajfasz J, Palmer SR, et al. Biology of Oral Streptococci. *Microbiol spectrum* 2018; 6.
36. Acuna-Castroviejo D, Escames G, Figueira JC, et al. Clinical trial to test the efficacy of melatonin in COVID-19. *J. Pineal Res* 2020; 69.
37. Aizawa S, Miyasawa-Hori H, Nakajo K, et al. Effects of alpha-amylase and its inhibitors on acid production from cooked starch by oral streptococci. *Caries res* 2009; 43:17-24.
38. Almughrabi OM, Marzouk KM, Hasanato RM, et al. Melatonin levels in periodontal health and disease. *J periodontal Res* 2013; 48:315-321
39. Anderson G, Reiter RJ. Melatonin: Roles in influenza, COVID-19, and other viral infections. *Rev Med Virol* 2020; 30.
40. Arastehfar A, Carvalho A, Nguyen MH, et al. COVID-19-Associated Candidiasis (CAC): An Underestimated Complication in the Absence of Immunological Predispositions? *J Fungi* 2020; 6:211.
41. Baghizadeh Fini M. Oral saliva and COVID-19. *Oral oncol* 2020; 108.
42. Barbieri DSV, Vicente VA, Fraiz FC, et al. Analysis of the *in vitro* adherence of *Streptococcus mutans* and *Candida albicans*. *Braz J Microbiol* 2007; 38:624-632.
43. Bencharit S, Altarawneh SK, Baxter SS, et al. Elucidating role of salivary proteins in denture stomatitis using a proteomic approach. *Mol Biosyst* 2020; 8:3216-3223.
44. Buranarom N, Komin O, Matangkasombut O. Hyposalivation, oral health, and *Candida* colonization in independent dentate elders. *PloS one* 2020; 15.
45. Caplan DJ, Hunt RJ. Salivary flow and risk of tooth loss in an elderly population. *Com Dent Oral Epidemiol* 1996; 24:68-71.
46. Castillo RR, Quizon GRA, Juco MJM, et al. Melatonin as adjuvant treatment for coronavirus disease 2019 pneumonia patients requiring hospitalization (MAC-19 PRO): a case series. *Melatonin Res* 2020; 3:297-310.
47. Cengiz MI, Cengiz S, Wang H. Melatonin and Oral Cavity. *Int J Dent* 2012.
48. Chen L, Zhao J, Peng J, et al. Detection of SARS-CoV-2 in saliva and characterization of oral symptoms in COVID-19 patients. *Cell Prolif* 2020; 53.
49. Claustrat B, Brun J, Chazot G. The basic physiology and pathophysiology of melatonin. *Sleep Med Rev* 2005; 9:11-24.
50. Culp DJ, Robinson B, Cash MN. Murine Salivary Amylase Protects Against *Streptococcus mutans*-Induced Caries. *Front physiol* 2021; 12.
51. Cutando A, Galindo P, Gomez-Moreno G, et al. Relationship between salivary melatonin and severity of periodontal disease. *J periodontal* 2006; 77:1533-1538.

52. Cutando A, Gomez-Moreno G, Arana C, et al. Melatonin: potential functions in the oral cavity. *J Periodontol* 2007; 78:1094-1102.
53. Dodds M, Roland S, Edgar M, et al. Saliva A review of its role in maintaining oral health and preventing dental disease. *BDJ Team* 2015; 2.
54. Farsi NM. Signs of oral dryness in relation to salivary flow rate, pH, buffering capacity and dry mouth complaints. *BMC oral health* 2007; 7:1-6:
55. Bacaksız F, Ebik B, Ekin N, et al. COVID-19 and Hyperamylasemia. *Authorea* 2021.
56. Gomez-Moreno G, Guardia J, Ferrera MJ, et al. Melatonin in diseases of the oral cavity. *Oral dis* 2010; 16: 242-247.
57. 33. Herrera D, Serrano J, Roldán S, et al. Is the oral cavity relevant in SARS-CoV-2 pandemic? *Clin Oral Invest* 2020; 24:2925-2930.
58. Kaabi SAG, MUSAFAER HK. Novel Phage Cocktail for the Treatment of Bacteria Causing Chronic Suppurative Otitis Media. *Trop J Nat Prod Res* 2020; 4:680-686.
59. Kalsbeek A, Buijs RM. Output pathways of the mammalian suprachiasmatic nucleus: coding circadian time by transmitter selection and specific targeting. *Cell Tissue Res* 2002;309:109-118.
60. Kitamura M, Kiyak HA, Mulligan K. Predictors of root caries in the elderly. *Community Dent Oral Epidemiol* 1986; 14:34-38.
61. Lai CC, Wang CY, Hsueh PR. Co-infections among patients with COVID-19: The need for combination therapy with non-anti-SARS-CoV-2 agents? *J Microbiol Immunol Infect Wei Mian Yu Gan Ran Za Zhi* 2020; 53:505-512.
62. Li X, Wang L, Yan S, et al. Clinical characteristics of 25 death cases with COVID-19: a retrospective review of medical records in a single medical center, Wuhan, China. *Int J Infect Dis* 2020; 94:128-132.
63. Li YC, Bai WZ, Hashikawa T. The neuroinvasive potential of SARS-CoV-2 may play a role in the respiratory failure of COVID-19 patients. *J Med Virol* 2020; 92:552-555.
64. Llana-Puy C. The role of saliva in maintaining oral health and as an aid to diagnosis. *Medicina oral, patologia oral y cirugía bucal* 2006; 11:449-455.
65. Lynge Pedersen AM, Belstrom D. The role of natural salivary defences in maintaining a healthy oral microbiota. *J dent* 2019; 80:3-12.
66. Marsh PD, Do T, Beighton D, et al. Influence of saliva on the oral microbiota. *Periodontol* 2000 2016; 70:80-92.
67. Mastrangelo A, Germinario BN, Ferrante M, et al. COVID-Bio Study Group. 2020. Candidemia in COVID-19 patients: incidence and characteristics in a prospective cohort compared to historical non-COVID-19 controls. *Clinical infectious diseases: an official publication of the Infectious Diseases Society of America*, ctaa 1594. Advance online publication.
68. Mortazavi H, Baharvand M, Movahhedian A, et al. Xerostomia due to systemic disease: A review of 20 conditions and mechanisms. *Annals Med Health Sci Res* 2014; 4:503-510.
69. Moser D, Biere K, Han B, et al. COVID-19 Impairs Immune Response to *Candida albicans*. *Front Immunol* 2021; 12:640-644.
70. Motahari P. Relationship of oral candidiasis with salivary lysozyme and lactoferrin in HIV-positive patients: a systematic review. *HIV and AIDS Review. Int J HIV-Related Problems* 2012; 20:17-20.
71. Murakami Y, Machino M, Fujisawa S. *Porphyromonas gingivalis* Fimbria-Induced Expression of Inflammatory Cytokines and Cyclooxygenase-2 in Mouse Macrophages and Its Inhibition by the Bioactive Compounds Fibronectin and Melatonin. *ISRN dent* 2012; 350-859.
72. Nambiar M, Varma SR, Jaber M, et al. Mycotic infections-mucormycosis and oral candidiasis associated with COVID-19: a significant and challenging association. *J oral microbiol* 2021; 13:1967699.
73. Noto Y, Sato T, Kudo M, et al. The relationship between salivary biomarkers and state-trait anxiety inventory score under mental arithmetic stress: a pilot study. *Anesthesia and Analgesia* 2005; 101:1873-1876.
74. Nowak R, McMillen IC, Redman J, et al. The correlation between serum and salivary melatonin concentrations and urinary 6-hydroxymelatonin sulphate excretion rates: two non-invasive techniques for monitoring human circadian rhythmicity. *Clin Endocrinol* 1987; 27:445-452.
75. Nowroozi N, Kawata T, Liu P, et al. High  $\beta$ -galactosidase and ganglioside GM1 levels in the human parotid gland. *Archives of Otolaryngology-Head Neck Surgery* 2001; 127:1381-1384.
76. Reiter RJ, Paredes SD, Manchester LC, et al. Reducing oxidative/nitrosative stress newly discovered gene for melatonin. *Criti Rev Biochemi Mol Biol* 2009;44: 175-200:
77. Romero AC, Ibuki FK, Nogueira FN. Sialic acid reduction in the saliva of streptozotocin induced diabetic rats. *Archiv Oral Biol* 2012; 57:1189-1193.
78. Saeralaathan S, Rajkumar A, Balaji TM, et al. Salivary melatonin is depleted in patients with dental caries due to the elevated oxidative stress. *J oral biol craniofac res* 2021; 11:547-551.
79. Salaric I, Karmelic I, Lovric J, et al. Salivary melatonin in oral squamous cell carcinoma patients. *Sci Rep* 2021; 11:13201.
80. Saldiva PHN, Braz-Silva PH, Caldini EG, et al. Salivary glands are a target for SARS-CoV-2: a source for saliva contamination. *J Pathol* 2021; 254:239-243.
81. Salehi M, Ahmadikia K, Mahmoudi S, et al. Oropharyngeal candidiasis in hospitalised

- COVID-19 patients from Iran: Species identification and antifungal susceptibility pattern. *Mycoses* 2020; 63:771-778.
82. Samaranyake YH, Samaranyake LP, Pow EH, et al. Antifungal effects of lysozyme and lactoferrin against genetically similar, sequential *Candida albicans* isolates from a human immunodeficiency virus-infected southern Chinese cohort. *J clin microbiol* 2001; 39:3296-3302.
83. Sardi JCO, Scorzoni L, Bernardi T, et al. *Candida* species: current epidemiology, pathogenicity, biofilm formation, natural antifungal products and new therapeutic options. *J med microbiol* 2013; 62:10-24.
84. Sebaa S, Hizette N, Boucherit-Otmani Z, et al. Dosedependent effect of lysozyme upon *Candida albicans* biofilm. *Molecular medicine reports* 2017; 15:1135-1142.
85. Sehirli A, Aksoy U, Koca-Unsal RB, et al. Role of NLRP3 inflammasome in COVID-19 and periodontitis: Possible protective effect of melatonin. *Med hypotheses* 2021; 151:110588.
86. Sharma A, Subramaniam P, Moiden S. Analysis of Salivary IgA, Amylase, Lactoferrin, and Lysozyme Before and After Comprehensive Dental Treatment in Children: A Prospective Study. *Contemporary Clin Dent* 2017; 8:526-530.
87. Shneider A, Kudriavtsev A, Vakhrusheva AV, et al. Can melatonin reduce the severity of COVID-19 pandemic? *Int Rev Immunol* 2020; 19:153-162.
88. Srinath R, Acharya AB, Thakur SL. Salivary and gingival crevicular fluid melatonin in periodontal health and disease. *J periodontol* 2010; 81:277-283.
89. Vaarala MH, Porvari KS, Kellokumpu S, et al. Expression of transmembrane serine protease TMPRSS2 in mouse and human tissues. *J Pathol* 2001;193:134-140.
90. Vakkuri O, Leppaluoto J, Kauppila A. Oral administration and distribution of melatonin in human serum, saliva and urine. *Life Sciences* 1985; 37:489-495.
91. Villa A, Connell CL, Abati S. Diagnosis and management of xerostomia and hyposalivation. *Ther Clin Risk Manag* 2014; 11:45-51.
92. Wang WK, Chen SY, Liu IJ, et al. SARS Research Group of the National Taiwan University/National Taiwan University Hospital. Detection of SARS-associated coronavirus in throat wash and saliva in early diagnosis. *Emerg Infect Dis* 2004; 10:1213-1219.
93. Xu X, Wang G, Ai L, et al. Melatonin suppresses TLR9-triggered proinflammatory cytokine production in macrophages by inhibiting ERK1/2 and AKT activation. *Sci Rep* 2018; 8:15579.
94. Xu J, Li Y, Gan F, et al. Salivary Glands: Potential Reservoirs for COVID-19 Asymptomatic Infection. *J dent res* 2020; 99-989.
95. Zhang R, Wang X, Ni L, et al. COVID-19: Melatonin as a potential adjuvant treatment. *Life Sci* 2020; 250:117583.
96. Zhou Y, Hou Y, Shen J, et al. Network-based drug repurposing for novel coronavirus 2019-nCoV/SARS-CoV-2. *Cell Discov* 2020; 6:1-18.